

**UNIVERSITY OF CATANIA**  
**INTERNATIONAL DOCTORAL PROGRAM IN**  
**ONCOLOGICAL SCIENCES**

**- CYCLE XXVIII -**

---

**Saverio Candido**

**Neutrophil gelatinase-associated lipocalin (NGAL) and  
matrix metalloproteinases (MMPs) as biomarkers of  
bladder cancer development and progression**

---

**DOCTORAL THESIS**

---

**Tutor: Prof. Massimo Libra**

**Coordinator: Prof. Ferdinando Nicoletti**

---

**ACADEMIC YEAR 2014/2015**

# CONTENTS

<b>ABSTRACT .....</b>	<b>1</b>
<b>1. INTRODUCTION .....</b>	<b>2</b>
1.1 NGAL.....	2
1.2 MMP-9.....	6
<b>2. RESULT .....</b>	<b>8</b>
2.1 COMPUTATIONAL ANALYSES OF NGAL IN DIFFERENT TUMOR TYPES .....	8
2.1.1 mRNA expression .....	8
2.1.2 Protein expression .....	12
2.1.3 Clinical impact .....	14
2.2 VALIDATION OF COMPUTATIONAL RESULTS IN BLADDER CANCER .....	26
<b>3. DISCUSSION .....</b>	<b>31</b>
3.4 FUTURE CHALLENGES .....	34
<b>4. MATERIAL AND METHODS.....</b>	<b>36</b>
4.1 COMPUTATIONAL ANALYSES.....	36
4.2 PATIENTS RECRUITMENT AND BLOOD SAMPLE COLLECTION .....	37
4.3 ELISA ASSAY .....	37
4.4 STATISTIC ANALYSIS .....	38
<b>5. REFERENCES .....</b>	<b>40</b>

## **ABSTRACT**

Neutrophil gelatinase-associated lipocalin (NGAL), also called lipocalin-2, is a secreted protein belonging to the lipocalin family proteins and actively participates into the proliferation, differentiation, and development of human tissues, including tumours. It positively modulates the activity of the matrix metalloproteinases-9 (MMP-9) that are involved in the enzymatic remodelling of the extracellular matrix. MMP-9 regulates the degradation of extracellular matrix in processes such as angiogenesis, tumour growth, and metastasis. By forming the NGAL/MMP-9 complex, NGAL protects MMP-9 from proteolytic degradation, a fundamental mechanism in controlling the activity of the proteins, and enhances its enzymatic activities.

As a secreted protein, NGAL is detectable in many biologic fluids, including urine, where several neoplastic cells and other tumor microenvironmental factors can be directly released from the cancer. Our *in silico* analysis suggested an active role of NGAL in tumour development of several cancer types. Validation of these findings is here described in bladder cancer as a good tumor model in which investigate the role of this protein because urine is in direct contact with the primary tumor.

On these bases, the release of NGAL in both urine and serum samples from 89 bladder cancer patients was measured. Further investigations, aimed to emphasize the role of NGAL in cancer, were performed by analysing MMP-9 and NGAL/MMP-9 complex levels in the same subset of bladder cancer patients. Control experiments were performed in 119 cancer-free controls, previously enrolled in a case-control study. Urinary concentrations were standardized on creatinine level. The performance of these proteins as cancer biomarkers was evaluated through the receiver operating characteristic (ROC) analysis.

In conclusion, the present study deepens the knowledge of the molecular mechanisms sustaining NGAL expression in tumor cells and its effects on cancer metastatic behaviour. Furthermore, NGAL/MMP-9 pathway is associated with an aggressive phenotype of transitional cell carcinoma of the bladder (TCC). The elevated negative predictive value of MMP-9 and NGAL/MMP-9 complex make them candidate markers of exclusion test for TCC. These findings suggest that these proteins may be integrated in the surveillance of bladder cancer, thus improving patients complaints and diminishing their discomfort.

## **1. INTRODUCTION**

Metastatic spreading is the major cause of death in cancer patients. Several molecules have been shown to contribute to tumor invasion and spreading. Recently, the role of neutrophil gelatinase-associated lipocalin (NGAL) has been associated with cancer development and progression [See Candido S et al, 2015 for a review]. However, its specific role for each tumor type was lacking and then better clarified in the present study.

NGAL is a secreted protein detectable in many biologic fluids, including urine, where several neoplastic cells and other tumor microenvironmental factors can be directly released into the urine. Therefore, bladder cancer may represent a good tumor model in which investigate the role of this candidate marker as urine is in direct contact with the primary tumor. Among tumor microenvironmental factors, matrix metalloproteinase-9 plays an important role, especially in tumor growth and spreading. Notably, NGAL's ability to combine in a dimeric complex with MMP-9, results in a protective action of MMP-9 from its auto-degradation and consequently results in a higher gelatinolytic action of MMP-9 on extracellular matrix [Yan L et al, 2001]. By this function, it has been shown that NGAL may promote cancer development in a variety of different cancer types [Fernández CA et al, 2005; Kubben FJ et al, 2007; Zhang H et al, 2007; Smith ER et al, 2008].

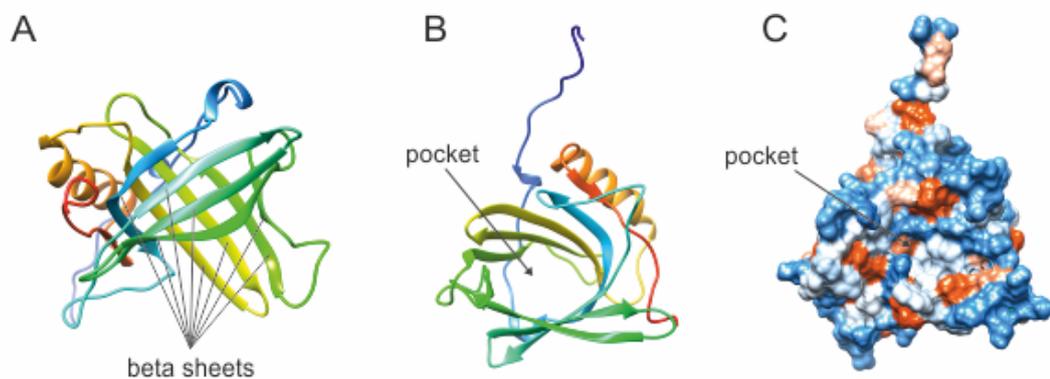
The analysis of NGAL transcript levels and its potential clinical implications in different cancer types, explored by bioinformatic approaches, represents the first part of the study included in this Ph.D. Program and it was already published in Oncotarget [See Candido S et al, 2014]. In detail, NGAL transcript levels were explored in different cancer types by analysing public available microarray datasets. NGAL protein expression were performed by analyzing the Human Protein Atlas. These computational data are thoroughly discussed below.

On these bases, validation of our computational data, representing the second part of the study included in this Ph.D. Program, was performed in a consecutive series of both blood and urine samples from bladder cancer patients and from age and sex-matched normal samples.

### **1.1 NGAL**

Members of the lipocalin protein family are characterized by their ability to bind small hydrophobic molecules (such as prostaglandins, retinoids, arachidonic acid, hormones and fatty acids). They often bind to specific cell-surface receptors and form macromolecular

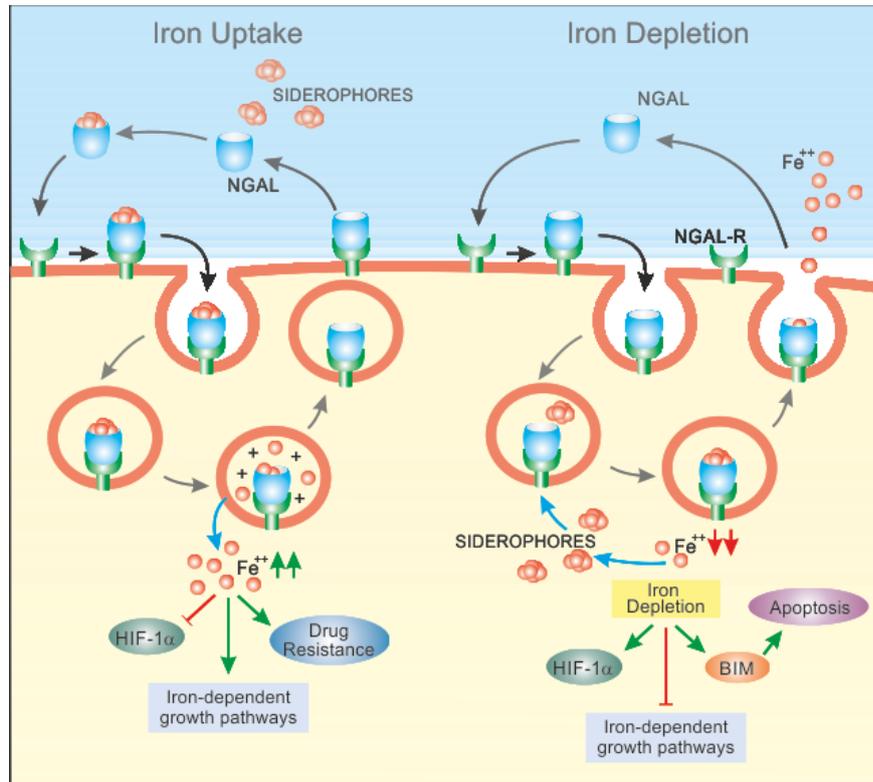
complexes. Highly conserved lipocalin crystal structures consist of a single eight-stranded continuously hydrogen-bonded antiparallel  $\beta$ -barrel delineating a calyx shape, which represents the internal ligand-binding site (Figure 1). Members of the lipocalin family, in the past classified exclusively as transport proteins, have now been described to carry out a variety of different functions. Some of these functions include: retinol transport, cryptic coloration, olfaction, pheromone transport, and the enzymatic synthesis of prostaglandins, moreover the lipocalins are also involved in the regulation of the immunoresponse and the mediation of cell homeostasis [Flower DR, 1996].



**Figure 1. Highly conserved lipocalin crystal structures consist of a single eight-stranded continuously hydrogen-bonded antiparallel  $\beta$ -barrel (A) delineating a calyx shape, which represents the internal ligand-binding site (B). Hydrophobicity surface (C).** Images were created from the RCSB PDB database (<http://www.rcsb.org>) (ID: 1NGL) using the UCSF Chimera package UFCS Chimera package that is developed by the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco (supported by NIGMS P41-GM103311). Ref: The solution structure and dynamics of human neutrophil gelatinase-associated lipocalin by Coles M, et al. J Mol Biol. 1999; 289: 139-57.

NGAL, also called lipocalin 2, siderocalin and 24p3, was identified in several forms: a monomer (25-kDa), a disulfide-linked homodimer (46-kDa), and a disulfide-linked heterodimer with human neutrophil gelatinase B (135-kDa) [Kjeldsen L et al, 1993]. NGAL has several functions. In early studies NGAL was described as a factor of innate immune system. NGAL is released by neutrophils at sites of infection and inflammation to sequester bacterial ferric siderophores, participating in the antibacterial iron-depletion strategy of innate immune system [Goetz DH et al, 2002]. Subsequently, it was shown that NGAL is responsible for iron delivery to the cytoplasm where it is accumulated and activates or represses iron-responsive genes. Iron unloading depends on the cycling of NGAL through acidic endosomes [Yang J et al, 2002]. In contrast, Devireddy LR et al have shown that NGAL is also involved in apoptosis-dependent deprivation of trophic factors. Apo-NGAL, after binding to its putative receptor, 24p3R, is internalized and associates with an intracellular siderophore, transferring chelated iron to the extracellular medium, thereby

reducing intracellular iron concentration which leads to the expression of the pro-apoptotic protein Bim, leading to the induction of apoptosis [Devireddy LR et al, 2005] (Figure 2).

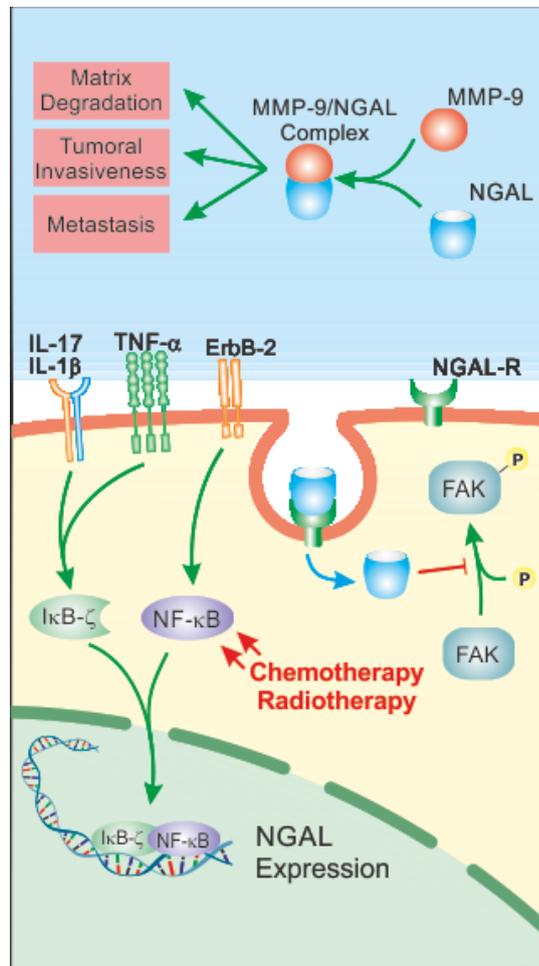


**Figure 2. Effects of iron concentration on NGAL activity.** NGAL in the presence of siderophores and ferrous iron ( $Fe^{++}$ ) is involved in iron uptake which is important in controlling the iron-dependent growth pathways, drug resistance and inhibiting the expression of HIF-1alpha. In contrast, under conditions of iron depletion, HIF-1alpha expression is stimulated, iron-dependent growth pathways are inhibited and apoptosis is induced. Abbreviations: Neutrophil gelatinase-associated lipocalin (NGAL), NGAL receptor (NGAL-R), hypoxia-inducible factor-1alpha (HIF-1-alpha), Bcl-2-like protein 11 (Bim).

NGAL was originally identified as a protein covalently associated with 92-kDa gelatinase/MMP9 from human activated neutrophils [Kjeldsen L et al, 1993]. NGAL is expressed in many other types of cells in response to various injuries, especially in kidney diseases. Serum NGAL levels correlate clearly with the severity of renal injury, reflecting the degree of tissue damage. For this reason, NGAL may become one of the most promising next-generation biomarkers in clinical nephrology and as well as other diseases and pathological states [Bolignano D et al, 2010].

NGAL is up-regulated by IL-1 beta, but not by TNF-alpha, in type II pneumocyte-derived cell line through the induction of the NF-kB pathway [Cowland JB et al, 2003]. IL-1 beta selectivity in inducing NGAL is due to the synthesis of I $\kappa$ B-zeta, a NF-kB-binding cofactor, elicited specifically by IL-1beta stimulation which is required for transcriptional activation of NGAL [Cowland JB et al, 2006]. Stimulation with TNF-alpha in the presence of IL-17, which

stabilizes the I $\kappa$ B-zeta transcript, is able to induce NGAL expression by I $\kappa$ B-zeta protein binding to NF- $\kappa$ B on the NGAL promoter [Karlsen JR et al, 2010]. It has been also demonstrated that activation of the NF- $\kappa$ B pathway is associated with up-regulation of NGAL-ErbB2-mediated signaling [Li SH et al, 2009] (Figure 3).



**Figure 3. Effects of cytokines and growth factor receptors on NGAL gene expression.**

Abbreviations: Matrix metalloproteinase 9 (MMP-9), v-Erb-B2 Avian Erythroblastic Leukemia Viral Oncogene Homolog (Erb-B2 = HER2), tumor necrosis factor-alpha (TNF- $\alpha$ ), interleukin-17 (IL-17), interleukin-1 beta (IL-1 $\beta$ ), focal adhesion kinase (FAK), nuclear factor kappa B (NF- $\kappa$ B) inhibitor of NF- $\kappa$ B zeta subunit (I $\kappa$ B $\zeta$ ), ferrous iron (Fe<sup>++</sup>).

Its complex with MMP-9 results in a higher gelatinolytic activity of MMP-9 on extracellular matrix [Yan L et al, 2001]. By this function, it was suggested that NGAL induce cancer development in several cancer types [Fernández CA et al, 2005; Kubben FJ et al, 2007; Zhang H et al, 2007; Smith ER et al, 2008]. Conversely, anticancer activities of NGAL have been demonstrated by its ability to inhibit the pro-neoplastic factor HIF-1 $\alpha$ , the synthesis of HIF-1 $\alpha$ -dependent VEGF [Venkatesha S et al, 2006; Tong Z et al, 2008], and phosphorylation of FAK kinase [Tong Z et al, 2008], as shown in colon [Lee HJ et al, 2006], ovarian [Lim R et

al, 2007] and pancreatic [Tong Z et al, 2008] cancers (Figure 3). Further evidences indicate that NGAL plays key roles in the inflammation and in the regulation of cell growth and adhesion in both normal and tumor tissues [Friedl A et al, 1999; Yang J et al, 2009; Bolignano D et al, 2010; Chakraborty S et al, 2012;].

## 1.2 MMP-9

Matrix metalloproteinase 9 (MMP-9) is a proteolytic enzyme belonging to the zinc-metalloproteinases family involved in the degradation of the extracellular matrix. MMP-9 was extensively studied in several cancer types, due to their dominant role in both tumor invasion and metastatic process [Shay G et al, 2015].

MMP-9 is involved in the degradation of extracellular matrix in normal physiological processes such as embryonic development, reproduction, angiogenesis, bone development, wound healing and cell migration [Vandooen J et al, 2013]. In these processes, the expression of MMP-9 is strictly regulated at transcriptional and post-transcriptional levels. Several factors such as cytokines, extracellular matrix proteins, cell/cell interactions and cell/matrix interactions are involved in MMP-9 transcriptional regulation, while at post-transductional level, the cleavage of MMP-9 proenzyme form (proMMP-9) is required to the full proteolytic activity [Sternlicht MD et al, 2001; Chen J et al, 2015] (Figure 4).

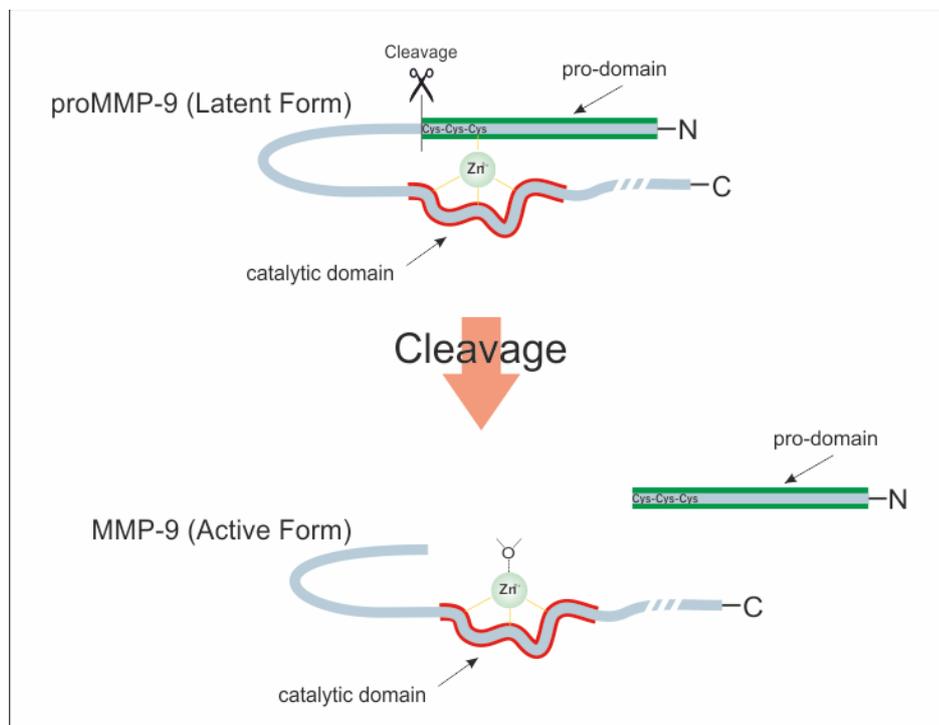


Figure 4. Proteolytic activation of proMMP-9.

In the presence of pathologic conditions, the expression of MMP-9 is up regulated by several inflammatory cytokines and growth factors that trigger the mitogen-activated protein kinases (MAPK) pathways leading to the trans-activation of the factors AP-1/PEA3 (Activator Protein-1/A Polyoma Enhancer Binding Protein-3), required for transcription of MMP-9 [Chen J et al, 2015]. In addition, MMP-9 transcription is regulated by the transcription factor NF-kB that is able to bind the MMP-9 promoter in response to inflammatory stimuli sustained by several factors, such as TNF- $\alpha$  [Shin SY et al, 2010].

Previous studies tried to determine which MMP-9 genetic modifications might be responsible of its upregulation. Among these, both polymorphisms and methylations of MMP-9 promoter have been considered [Yan C et al, 2007; Kulis M et al, 2013].

## **2. RESULT**

### **2.1 COMPUTATIONAL ANALYSES OF NGAL IN DIFFERENT TUMOR TYPES**

#### **2.1.1 mRNA expression**

Gene expression patterns of NGAL mRNAs present in different tumor types were obtained from several datasets (Table 1). Significant differences between tumor tissues and relative normal counterparts for each cancer type are reported in Table 1.

Significant differences between tumor tissues and relative normal counterparts for each cancer type are reported in Table 1. This analysis showed that NGAL transcript levels were significantly higher in the majority of solid tumors compared to the relative normal tissues for every dataset analyzed. While, lower levels of NGAL were observed in each dataset of cervical cancer, esophageal cancer, head and neck cancer and in haematological malignancies. In Table 2 are presented the significant differences of NGAL transcript levels observed in metastatic tissues compared to those of the relative primary tumor. The results showed that in all dataset analyzed the levels were significantly lower in the metastatic tissues than in primary tumors from 5 different tumor types, including colorectal, kidney, melanoma, ovarian and prostate (Table 2).

In Figure 5 the distribution of NGAL transcript levels among cancer cases and normal samples is shown. The percentage of tumor cases showing NGAL transcript levels below the 25th percentile and above the 75th percentile of the “normal” samples is also reported.

**Table 1.** Gene expression patterns of NGAL in different cancer types from 29 datasets

Cancer Type	# of samples		Fold Change ( $\leq -2$ ) or ( $\geq 2$ )	p < 0.01 (T-test)	Dataset		
	Cancer	Normal			Author (ref.)	Year	Platform
<b>Solid tumor</b>							
Bladder	109	48	4.13	2.75E-05	Sanchez-Carbayo M <i>et al</i>	2006	U133A
Cervix	32 <sup>a</sup>	24	-3.21	4.06E-04	Scotto L <i>et al</i>	2008	U133A
Colon	95	5	2.62	1.23E-02	Kaiser S <i>et al</i>	2007	U133 Plus 2.0
	81	24	4.15	7.04E-08	Skrzypczak M <i>et al</i>	2010	U133 Plus 2.0
	70	12	4.78	7.06E-06	Hong Y <i>et al</i>	2010	U133 Plus 2.0
Esophagus	17 <sup>b</sup>	17	-5.41	9.27E-05	Hu N <i>et al</i>	2010	U133A
	53 <sup>b</sup>	53	-2.92	1.05E-06	Su H <i>et al</i>	2011	U133A/B
Head and Neck	6 <sup>d</sup>	4	-18.58	7.33E-03	Schlingemann J <i>et al</i>	2005	U133A
	31 <sup>c</sup>	10	-12.44	3.60E-09	Sengupta S <i>et al</i>	2006	U133 Plus 2.0
kidney	51 <sup>e</sup>	5	4.15	4.98E-05	Yusenko MV <i>et al</i>	2009	U133 Plus 2.0
Liver	35	10	8.66	2.32E-05	Wurmbach E <i>et al</i>	2007	U133 Plus 2.0
	22	21	3.48	5.14E-04	Roessler S <i>et al</i> (1)	2010	U133 Plus 2.0
	225	220	2.94	6.86E-22	Roessler S <i>et al</i> (2)	2010	HT U133A
Lung	30 <sup>f</sup>	30	2.79	1.72E-03	Su LJ <i>et al</i>	2007	U133A
	58 <sup>f</sup>	49	2.28	2.64E-06	Landi MT <i>et al</i>	2008	U133A
	226 <sup>f</sup>	20	3.707	1.53E-7	Okayama H <i>et al</i>	2012	U133 Plus 2.0
Ovary	185 <sup>g</sup>	10	5.84	1.54E-06	Bonome T <i>et al</i>	2008	U133A
	99 <sup>g</sup>	4	3.03	6.98E-04	Hendrix ND <i>et al</i>	2006	U133A
Pancreas	36 <sup>h</sup>	16	14.05	5.15E-06	Pei H <i>et al</i>	2009	U133 Plus 2.0
	11 <sup>h</sup>	6	10.00	9.03E-05	Segara D <i>et al</i>	2005	U133A
	39 <sup>i</sup>	39	7.70	1.64E-10	Badea L <i>et al</i>	2008	U133 Plus 2.0
Thyroid	9 <sup>j</sup>	9	3.77	9.76E-4	He H <i>et al</i>	2005	U133 Plus 2.0
	14 <sup>j</sup>	4	2.33	0.001	Vasko V <i>et al</i>	2007	U133 Plus 2.0
	26 <sup>j</sup>	4	2.05	9.34E-7	Giordano TJ <i>et al</i>	2006	U133A
<b>Hematologic tumor</b>							
ALL	750	74	-11.77	7.57E-152	Haferlach T <i>et al</i>	2010	U133 Plus 2.0
AML	542	74	-16.94	2.32E-165	Haferlach T <i>et al</i>	2010	U133 Plus 2.0
	285	8	-4.91	5.00E-03	Valk PJ <i>et al</i>	2004	U133A
CLL	448	74	-42.98	7.73E-194	Haferlach T <i>et al</i>	2010	U133 Plus 2.0
Myeloma	9 <sup>k</sup>	5	-3.90	0.002	Agnelli L <i>et al</i>	2009	U133A

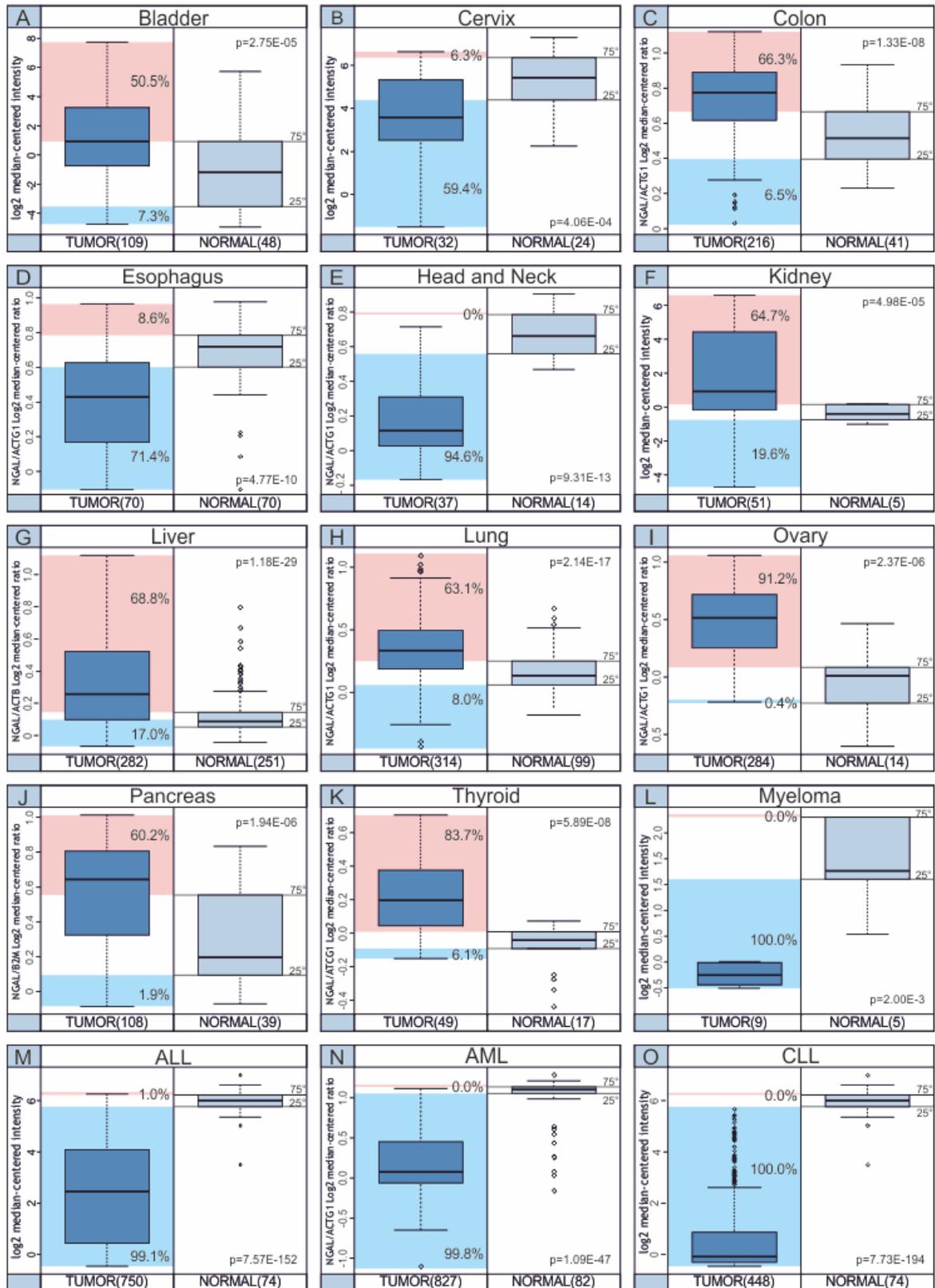
**Legend: Solid tumor:** Cervix: <sup>a</sup>Cervical Squamous Cell Carcinoma. Esophagus: <sup>b</sup>Esophageal Squamous Cell Carcinoma; Head and Neck: <sup>c</sup>Nasopharyngeal Carcinoma; <sup>d</sup>Squamous Cell Carcinoma. Kidney: <sup>e</sup>Renal Carcinoma; Liver: (1), dataset 1; (2) dataset 2; Lung: <sup>f</sup>Lung adenocarcinoma; Ovary: <sup>g</sup>Ovarian Carcinoma; Pancreas: <sup>h</sup>Pancreatic Carcinoma; <sup>i</sup>Pancreatic Ductal Adenocarcinoma; Thyroid: <sup>j</sup>Thyroid Gland Papillary Carcinoma.

**Hematologic tumor:** ALL, Acute Lymphoblastic Leukemia; AML, Acute Myeloid Leukemia; CLL, Chronic Lymphocytic Leukemia. Myeloma: <sup>k</sup>Plasma Cell Leukemia.

**Table 2.** NGAL transcripts in metastatic tissues compared to the relative primary tumor

Cancer Type	N. of samples		FC ( $\leq -1.5$ ) or ( $\geq 1.5$ )	p < 0,05 (T-test)	Data set		
	metastasis	primary			Author (ref.)	Year	Platform
Colorectal	43	330	-3.191	1.56E-06	Bittner M [a]	2005	U133 Plus 2.0
	27	56	-5.437	1.04E-06	Tsuji S <i>et al</i>	2012	U133 Plus 2.0
Kidney	60	9	-1.960	4.00E-03	Jones J <i>et al</i>	2005	U133A
Melanoma	40	16	-2.475	3.00E-03	Riker AI <i>et al</i>	2007	U133 Plus 2.0
	52	31	-4.849	1.12E-08	Xu L <i>et al</i>	2008	U133A
Ovarian	75	166	-1.999	2.00E-03	Bittner M [a]	2005	U133 Plus 2.0
	16	74	-1.537	4.30E-03	Anglesio MS <i>et al</i>	2008	U133 Plus 2.0
Prostate	5	27	-1.578	2.60E-02	Vanaja DK <i>et al</i>	2003	U133A/B
	6	7	-5.735	7.76E-04	Varambally S <i>et al</i>	2005	U133 Plus 2.0

[a] GEO Series GSE2109; FC, Fold change.



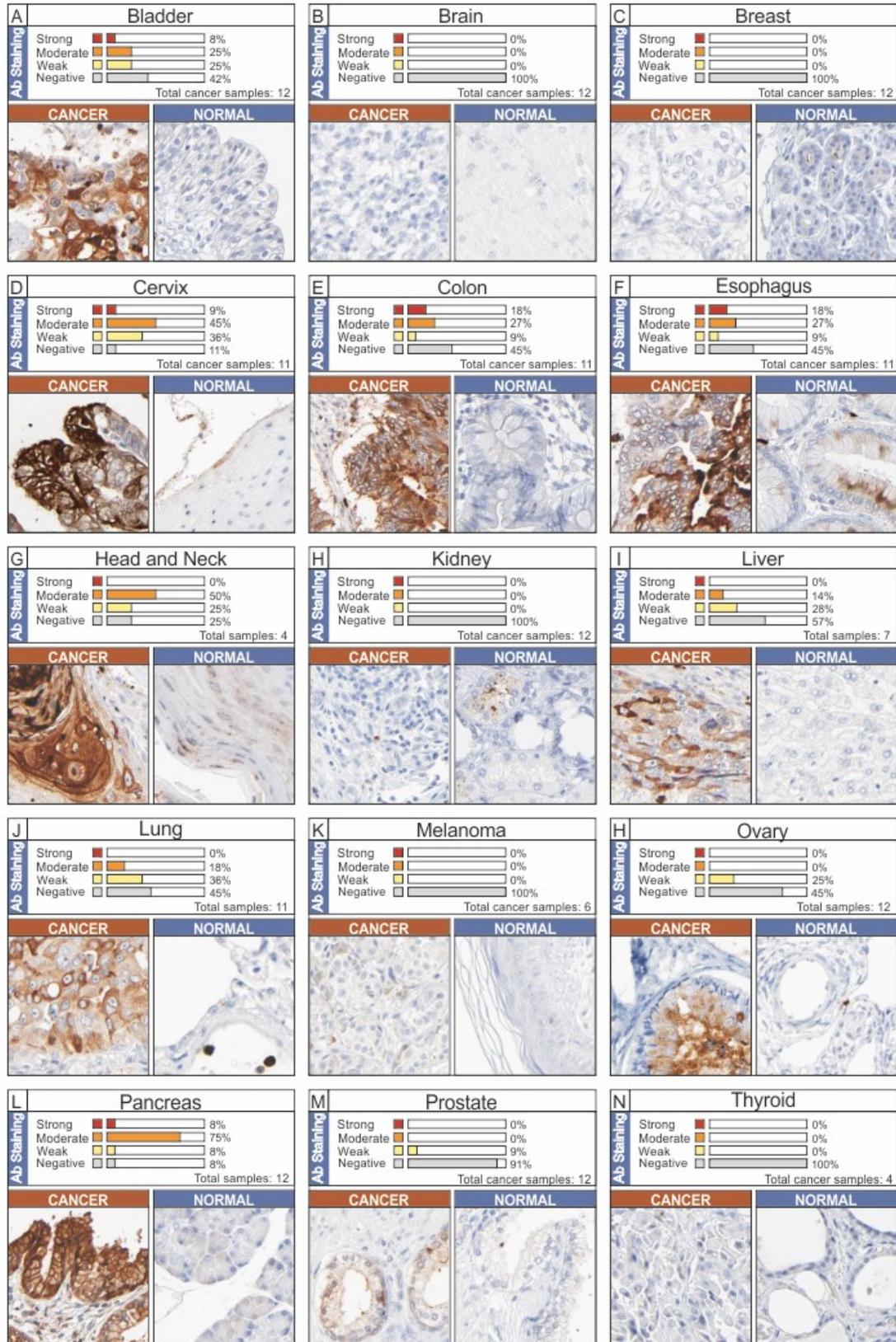
**Figure 5. Distribution of NGAL transcript levels among cancer cases and normal samples.** The percentage of tumor cases, indicated for each tumor setting, shows NGAL transcript levels below the 25<sup>th</sup> percentile (Cyan box) and above the 75<sup>th</sup> percentile (Magenta box) of the “normal” samples.

### **2.1.2 Protein expression**

To understand if there was an association between mRNA and protein NGAL expression, an immunohistochemistry evaluation was performed by analysis of Human Protein Atlas web site. In Figure 6 immunohistochemistry analysis of NGAL in 15 solid cancer types are shown. Cancer types with negative immunostaining are not shown. The data demonstrated that concordance of NGAL at both mRNA and protein levels was obtained for the following cancer types: bladder, colorectal, liver, lung, ovarian, and pancreatic (Table 1 and Figure 6 Panels: A, E, I, J, H and L). Cervical, esophageal/stomach and head and neck cancers showed a moderate positive immunostaining (Figure 6, panel D, F and G), while mRNA expression levels were lower in cancer tissue compared to the normal counterpart (Table 1).

Conversely, weak immunostaining (<10% of cases) or negative protein expression levels were observed in brain, breast, melanoma and prostate cancers. Accordingly, no significant differences in NGAL mRNA expression were observed between cancer and relative normal counterpart in all datasets analysed for each of these 4 cancer types (brain, breast, melanoma and prostate) (Figure 6, Panel B, C, K and M).

Although, higher mRNA levels were detected in renal and thyroid cancers than in relative normal counterparts (Table 1), a negative immunostaining was observed (Figure 6, Panel H and N). Finally, concordant negative mRNA and protein levels of NGAL were observed in the majority of haematological malignancies (data not shown).



**Figure 6. Immunohistochemistry analysis of NGAL expression in human cancer.** The data were obtained from the Human Protein Atlas. A single representative case for each cancer type (total 15) is shown along with its normal counterpart. Expression of NGAL in cancer sample was evaluated as strong, moderate, weak and negative immunostaining. The percentage is referred to the total cancer samples analyzed for each tumor type.

### **2.1.3 Clinical impact**

The role of NGAL for each tumor type is described below and summarized in Table 3.

#### **SOLID TUMORS**

##### **Bladder**

The current study reveals that both mRNA transcript and protein levels were higher in bladder cancer tissues than in the normal counterparts. Accordingly, 50.5% of cases displayed NGAL transcripts above the 75th percentile of the “normal” values suggesting its role as a diagnostic marker (Figure 5A). These data are in agreement with previous investigations in which NGAL along with MMP-9 were overexpressed in urothelial bladder carcinomas suggesting their role as early diagnostic markers for this tumor type [Roy R et al, 2008; Monier F et al, 2000]. Reduced protein levels of both NGAL and MMP-9 have been detected in urine samples from bladder cancer patients with clinical relapse suggesting that reduced levels of these proteins may be used as indicators of tumor progression [Monier F, et al 2002].

##### **Brain**

A significant association of MMPs and NGAL expression was detected in urine from brain cancer patients and in tumor specimens. Additionally, surgical resection of the tumor resulted in a reduction of both MMPs and NGAL in urine samples [Smith ER et al, 2008]. The identification of new potential molecular targets in this cancer type may be very helpful to discover new therapeutic strategies. Immunohistochemistry analysis reveals that NGAL expression is frequently up-regulated in gliomas and is associated with poor clinical outcome [Liu MF et al, 2011]. Barresi et al show that NGAL was overexpressed in primary high grade brain tumors and not in the metastatic cases [Barresi V et al, 2010 (4)]. According to the exclusion criteria designed for the purpose of our present study, no data on brain cancer were generated by computational analysis. Accordingly, NGAL protein expression was not detected through the Human Protein Atlas evaluation.

##### **Breast**

Similar observations acquired for brain tumors were obtained for breast cancer as no data were generated by our computational analysis. The expression pattern of NGAL has been evaluated by several authors and generated conflicting results on the clinical significance of this protein in breast cancer.

A heterogeneous pattern of expression at the mRNA and protein levels was observed in breast cancer patients in a study conducted by Stoesz SP et al [Stoesz SP et al 1998]. The authors also described a significant correlation between NGAL expression and other markers of poor prognosis, including estrogen and progesterone receptor-negative status and high proliferation (S-phase fraction) [Stoesz SP et al 1998]. These studies were also confirmed by Bauer M et al [Bauer M et al, 2008]. Similarly, Shen ZZ et al showed that MMP-9 and MMP-9/NGAL complex expression were higher in breast cancer than in benign breast and/or normal tissues [Shen ZZ et al, 2003]. While, in the study conducted by Provatopoulou X et al. in a large series of breast cancer patients, MMP-9 and NGAL were overexpressed in cancer and this overexpression was associated with the severity of disease but no significant correlation was found for the complex formation [Provatopoulou X et al 2009].

In a transgenic mouse model of breast cancer, Berger et al. demonstrated that lack of NGAL in mice leads to a reduction of tumor growth. This reduction was attributed to an NGAL-dependent decrease of MMP-9 activity and to a lack of high molecular weight MMP activity [Berger T et al, 2010]. Accordingly, Li et al have shown that NGAL expression is associated with increased metastasis and poor prognosis in breast cancer patients [Li SH et al 2009]. The results obtained by Leng et al in an animal model of breast cancer suggest that the suppression of NGAL function, by an inhibitory monoclonal antibody, has a great potential for breast cancer therapy, particularly by interfering with metastasis in aggressive types of breast cancer [Leng X et al, 2009]. Conversely, in a previous study it was shown that NGAL overexpression promotes in vivo the development of lung metastasis [Shi H et al, 2008]. Our recent studies indicate that increased NGAL expression did not alter the sensitivity of the MCF-7 breast cancer cell line to the chemotherapeutic drug doxorubicin [Chappell WH et al, 2012 (1)]. However, ectopic NGAL expression did alter the sensitivity of breast cancer cells to targeted therapy [Chappell WH et al, 2012 (2)]. Furthermore, NGAL was found to be a predictive marker for complete response after neo-adjuvant chemotherapy in low-risk subgroups of breast cancer patients and may be considered as an independent prognostic factor for decreased disease free survival in primary human breast cancer [Wenners AS et al, 2012].

## **Cervical cancer**

A recent analysis conducted by immunohistochemistry on a set of cervical biopsy specimens from 225 women showed a close relationship between NGAL expression levels, HPV lesion grade and detection of high risk HPV types. Up-regulation of NGAL in higher grade lesions is likely to be from the suppression of wild-type p53 by the HPV E6 oncoprotein. Suppression of p53 results in elimination of p53 block of NGAL transcription [Syrjänen S et al, 2010]. These data were in agreement with our IHC evaluation (Figure 6D) but not in line with our computational analysis as only 6.3% of cervical cases displayed NGAL transcripts above the 75th percentile of “normal” values, while 59.4% below the 25th percentile (Figure 5B).

## **Colorectal cancer**

In 1996 Nielsen et al analyzed the role of NGAL by both immunohistochemistry and mRNA by in situ hybridization in colon cancer and in inflammatory colorectal diseases. Increased expression of NGAL was detected both in non-malignant epithelium such as diverticulitis, inflammatory bowel disease and in malignant colonic lesions. In adenocarcinomas, NGAL overexpression was observed in the transitional mucosa and in the superficial ulcerated area. On the other hand, NGAL expression was not detected in lymph node metastases from this adenocarcinoma [Nielsen BS et al, 1996]. The authors speculate that NGAL is predominantly involved in inflammatory reaction and in tumor transformation, while it does not appear to play a prominent role in metastatic process.

However, Lee et al have shown that NGAL may function as a metastasis suppressor in colon cancer cells. In their studies, they genetically manipulated highly metastatic human colon cancer cell lines, which normally express low NGAL protein levels, to overexpress NGAL. Ectopic expression of NGAL suppressed, in vivo, the liver metastasis of metastatic human colon cancer cell lines in experimentally-driven metastasis assays [Lee HJ et al, 2006]. Hu et al examined the potential molecular mechanism of NGAL involvement in colorectal cancer. They demonstrated that NGAL overexpression altered the subcellular localization of E-cadherin and catenins, decreased E-cadherin-mediated cell to cell adhesion, enhanced cell-matrix attachment, and increased cell motility and in vitro invasion. They proposed that NGAL exerted these effects through the alteration of the subcellular localization of Rac1, one of Rho small GTPases, in an extracellular matrix-dependent manner, but not by MMP-9 [Hu L et al, 2009]. Recently, Bousserouel S et al have shown, in a preclinical model of colon

carcinogenesis, that NGAL is significantly upregulated only in advanced stages of tumor progression [Bousserouel S et al, 2010].

Real time PCR and zymographic analysis on visceral adipose tissue (VAT) biopsies from 11 colon cancer patients revealed increased levels of NGAL and other inflammation associated factors like osteopontin, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), and chitinase-3 like-1 compared to control subjects, suggesting their involvement in cancer development and progression [Catalán V et al, 2011].

Recently, Martí J et al showed the prognostic utility of NGAL mainly in metastatic CRC [Martí J et al, 2010; Martí J et al 2013]. Higher levels in colon cancer cases than controls were observed by Fung KY et al but the authors concluded that it was not a promising biomarker for the diagnosis of CRC as the sensitivity of NGAL was found to be 24% at 90% specificity [Fung KY et al, 2013]. Accordingly, although NGAL is still expressed by the majority of human neoplastic colorectal lesions, the author it is not a useful biomarker for determining disease progression from adenomatous to malignant colorectal neoplasia [McLean MH et al, 2013]. Our analysis, in line with the majority of previous studies, shows the upregulation of NGAL in adenocarcinoma tumor samples and reduced expression in metastatic samples.

### **Esophageal cancer**

Esophageal cancer is the eighth most common incident cancer in the world and ranks sixth among all cancers in mortality. Esophageal cancers are classified into two histological types; esophageal squamous cell carcinoma (ESCC), and adenocarcinoma, and the incidences of these types show a striking variety of geographic distribution, possibly reflecting differences in exposure to specific environmental factors. Both alcohol consumption and cigarette smoking are major risk factors for the development of ESCC [Koshy M et al, 2004].

Zhang et al performed immunohistochemistry, western blot and gelatin zymography on 81 paraffin sections including ESCC, normal mucosa, simple hyperplasia and dysplasia, and on 73 fresh specimens of ESCC to evaluate the role of NGAL in ESCC. Immunohistochemical studies revealed that ESCC have a diverse and obvious whole-cytoplasmic staining pattern for NGAL, while normal esophageal epithelium presented a weak positive signal within a restricted cytoplasmic area. On western blot analysis, NGAL expression level was found to be significantly higher in ESCC than in normal mucosa, and positively correlated with cancer cell differentiation. No significant association was observed between NGAL expression and cell proliferation. Finally, the authors showed higher enzymatic activity of MMP-9/NGAL

complex in ESCC than in normal mucosa. These findings suggest that NGAL is involved in differentiation pathways and invasive progression of ESCC [Zhang H et al, 2007]. Similar data were reached by Du et al, [Du ZP et al, 2011]. Accordingly our IHC evaluations show that NGAL is overexpressed in tumor and not in normal tissue. However, different trend is observed by analyzing the transcript levels as 71.4% of cases displayed NGAL transcripts below the 25th percentile of “normal” values, while 8.6% above the 75th percentile (Figure 5D).

Fang et al. identified a new NGALR isoform designated as NGALR-3, that results from alternative splicing. Interestingly, it was shown that the NGALR-3 isoform was overexpressed in 70% of esophageal carcinoma cases in comparison with those of normal adjacent epithelium. These findings suggest that the new NGALR-3 variant could play a more important role in esophageal carcinoma. The authors proposed that the novel NGALR-3 isoform could mediate a unilateral intracellular iron-delivery pathway which increased intracellular iron levels. This could be involved in the tumor growth of esophageal carcinomas [Fang WK et al, 2007].

Li EM et al studied the role of NGAL in invasion, division and proliferation of an esophageal carcinoma cell line. Their results demonstrated that the antisense blocking of NGAL transcription not only decreased effectively the activity of MMP-9 and MMP-2 secreted by SHEEC cells, but also suppressed significantly the invasion of these cells in nude mice. However, it was shown that NGAL was not apparently related with division and proliferation of SHEEC tumor cells [Li EM et al, 2003 (1)]. Furthermore, the same authors, in attempt to demonstrate the regulation mechanism of NGAL overexpression in SHEEC, analyzed the structural characters of 5'-untranslated region (5'-UTR) and 3'untranslated region (3'-UTR) of NGAL. Upon cloning and DNA sequencing of 69 bp 5'-UTR and 147 bp 3'-UTR of NGAL gene they did not observe any base pair mutations [Li EM et al, 2003 (2)].

### **Gastric cancer**

Gastric cancer (GC) is the final result of a multistep process initiated by environmental factors, including diet and Helicobacter pylori infection [Stemmermann GN et al, 2002]. H. pylori infection is one of the most important risk factors for this malignancy [Uemura N et al, 2001; Huang JQ, 2003].

During infection, H. pylori synthesize siderophores, which chelate Fe<sup>3+</sup> with high affinity and facilitate its transport into the pathogen [Neilands JB, 1995; Braun V et al, 2002;

Krewulak KD et al, 2008]. The host cells respond to infection by increasing the secretion of NGAL that binds the bacterial siderophores and prevents their uptake into bacteria. The iron depletion results in inhibiting *H. pylori* growth [Holmes MA et al, 2005].

NGAL and MMP9/NGAL complex were shown to be upregulated in GC tissue (mainly in neutrophils and epithelial cells) compared to adjacent normal gastric mucosa, confirming the hypothesis that the association of NGAL with MMP9 could prevent extracellular autodegradation of the proteinase. Enhanced levels of the MMP9/NGAL complex, but not of MMP-9 and NGAL, have been related to worse clinical outcome in cancer patients and significantly associated with the classifications of Lauren and WHO, suggesting that MMP9/NGAL complex could be considered as a novel prognostic factor for gastric cancer [Kubben FJ et al, 2007].

Wang HJ et al [Wang HJ et al, 2010] proposed NGAL as a potential biomarker for prognosis and an ancillary diagnostic test of gastric cancer. In this study, they showed high levels of NGAL expression in 333 GC patients by immunohistochemistry. NGAL was correlated with size of tumor, Lauren's classification, lymph node metastasis, vascular invasion, distant metastasis and TNM stage. The multivariate analysis indicated that NGAL can be used as an independent prognostic factor. Serum NGAL levels were determined in blood samples from 63 healthy donors and 60 GC patients and analyzed according to TNM. NGAL blood levels were higher than those of CA19-9 in TNM I patients, and higher than those of CEA and CA19-9 in TNM II. Therefore, serum NGAL has a great potential as a tumor marker for GC and could be associated with a poor prognosis [Wang HJ et al, 2010].

According to the exclusion criteria designed for the purpose of our present study, no data on GC were generated by our analyses.

### **Head and neck**

Our “in silico” analysis showed a significant down-regulation of NGAL in head and neck cancer compared to the normal counterpart. In fact, 94.6% of head and neck cancer displayed NGAL transcripts below the 25th percentile of “normal” values, while no samples showed mRNA levels of NGAL above the 75th percentile (Figure 5E). However, our IHC evaluation is in line with previous data [Lin CW et al, 2012] as expression of NGAL appeared to be moderate in 50% of cases and absent in normal tissue.

## **Hepatocellular Carcinoma (HCC)**

Zhang Y et al demonstrated the up-regulation of NGAL expression in HCC was significantly correlated with unfavorable clinic-pathologic features and independent poor prognostic factor for overall survival in patients [Zhang Y et al, 2012]. In agreement, our analysis showed a significant up-regulation of NGAL mRNA levels in HCC from 2.3- to 8.6-fold changes in comparison to normal hepatic tissues in the three datasets analyzed (Table 1). Similar data were obtained by IHC considerations.

## **Lung cancer**

At the present, few studies have been performed to evaluate the role of NGAL in lung cancer. However, Friedl A et al found high NGAL levels in lung adenocarcinoma [Friedl A et al, 1999]. Accordingly, in the present study we detected a constant NGAL upregulation both at mRNA and protein levels for adenocarcinoma histotype. In contrast, NGAL was not overexpressed in carcinoid lung cancer.

## **Melanoma**

In melanoma a substantial down-regulation of NGAL was observed only in metastatic disease versus primary tumor while no statistic differences were observed in NGAL mRNA levels between primary tumor and normal tissue. Protein expression of NGAL was not detected by IHC (Human Protein Atlas web site) (Figure 6K). To our knowledge no previous data were generated on NGAL in melanoma tissue samples.

## **Ovarian cancer**

Epithelial ovarian cancer is one of the most aggressive cancers diagnosed in women with high mortality rate. This cancer is usually asymptomatic and often diagnosed late in the disease process [Paley PJ, 2001]. Unfortunately, there are no specific markers for early diagnosis. Lim R et al analyzed NGAL by IHC in a total of 59 ovarian tissues including normal, benign, borderline and malignant (grades 1, 2 and 3). NGAL expression was weak or moderate in benign tissues. Both borderline and grade 1 tumors displayed the highest amount of NGAL expression with moderate to strong staining, whereas in grade 2 and 3 tumors, the extent of staining was significantly less ( $p < 0.01$ ) and staining intensity was weak to moderate. Additionally, the authors analyzed, by ELISA, NGAL levels in 62 serum samples from normal individuals and ovarian cancer patients (grade 1). The NGAL concentration was 2 and

2.6-fold higher in patients with benign tumors and cancer patients (grade 1) [Lim R et al, 2007]. In line with results provided by Lim et al, all datasets here examined showed an upregulation of NGAL in ovarian cancer samples, with a range from 3 to 6 fold. Furthermore, the analysis of both Bittner and Anglesio datasets (Ref. in Table 2) revealed a significant reduction of NGAL mRNA levels in metastatic disease (see Table 2). In the same study, Lim and collaborators analyzed NGAL in ovarian cancer cell lines treated with epidermal growth factor (EGF) indicating that NGAL expression was downregulated in ovarian cancer cell lines undergoing the epithelial to mesenchymal transition (EMT) induced by EGF. Downregulation of NGAL expression correlated with the upregulation of vimentin, enhanced cell dispersion and downregulation of E-cadherin expression, some of the hallmarks of EMT. These data indicate that NGAL may be a good marker to monitor transformation of benign lesions to premalignant and malignant ovarian tumors and that the molecule may be involved in the progression of epithelial ovarian malignancies [Lim R et al, 2007].

NGAL, as well as other proteins, was shown to be regulated in ovarian cancer cell lines by 17 beta-estradiol/estrogen [Walker G et al, 2007]. More recently Cho et al. described the upregulation of NGAL in a panel of 54 ovarian cancers, 15 borderline and 53 benign ovarian tumors, and 90 healthy controls by real time PCR and immunohistochemical analysis. NGAL levels were significantly higher in ovarian tissues and particularly in well-differentiated tumors. Similar results were obtained by analyzing NGAL serum levels from ovarian cancer patients showing highest levels in differentiated cancer [Cho H et al, 2009]. Notably, according to previous data, the results of the present study show that 91,2% of ovarian cases displayed NGAL transcripts above the 75th percentile of the “normal” values, while 0.4% below the 25th percentile. Similar trend was obtained for the IHC analyses. While transcript levels of NGAL were significantly lower in metastatic setting compared to primary tumor.

### **Pancreatic cancer**

Pancreatic cancer (PaCa) is the fifth leading cause of cancer death in both men and women [Ahmedin J et al, 2002]. Early detection of this disease is not possible in spite of significant diagnostic tools. The effective therapy, surgery, is limited to about 25% of the cases and often is unable to prevent cancer recurrence in these patients [Cooperman AM, 2001]. Much remains to be understood about the natural course and biology of this disease.

Using microarray analysis, many laboratories have reported the differential expression of several novel genes, including that of NGAL, associated with the progression of pancreatic

cancer [Argani P et al, 2001; Han H et al, 2002; Terris B et al, 2002]. Moniaux et al detected NGAL levels by immunohistochemistry on tissue samples from normal patients, pancreatitis, and pancreatic adenocarcinoma patients. Their results revealed higher levels in pancreatic adenocarcinoma than in normal and pancreatitis samples [Moniaux N et al, 2008]. These findings are in agreement with the results of the present study in which we observe a constant upregulation of NGAL at both mRNA and protein expression levels in pancreatic adenocarcinoma samples vs normal tissues. Moniaux et al also evaluated NGAL levels in pancreatic cancer cell lines with varying grades of differentiation documenting a positivity for NGAL expression in both well and moderately differentiated cells. In contrast, NGAL expression was uniformly negative in poorly differentiated adenocarcinoma. Further, they examined NGAL levels in serum samples. They used ELISA to detect NGAL. The authors concluded that NGAL was fairly accurate in distinguishing between pancreatic cancers and non-cancer cases. In conclusion, NGAL is highly expressed in early pancreatic dysplastic lesions, suggesting a possible role as an early diagnostic marker for pancreatic cancer [Moniaux N et al, 2008].

NGAL is also detected in bile and may be useful as a novel biomarker to distinguish benign from malignant biliary obstruction [Zabron AA et al, 2011].

The biological roles of NGAL in pancreatic adenocarcinoma have been studied both in vitro and in vivo by Tong et al [Tong Z et al, 2008]. The authors transfected PaCa cells with NGAL and demonstrated no effects on cell viability or sensitivity to chemotherapy, but decreases in cell adhesion, invasion and angiogenesis was observed both in vitro and in vivo. The negative effects on tumor progression and metastasis were due to alteration of FAK phosphorylation and decrease of VEGF production [Tong Z et al, 2008]. We can gather from this study that modulation of NGAL activity could control PaCa angiogenesis and metastasis.

### **Prostate cancer**

According to the exclusion criteria designed for the purpose of our present study, no differences were identified between prostate cancer tissue and the normal counterparts. Similarly, IHC evaluation did not display any differences between normal and tumor. While, significant differences were observed in metastatic disease of prostate cancer when compared with primary tumors as NGAL transcripts were lower in metastatic setting compared to primary tumor. However, in vitro studies showed that NGAL plays a significant role in the progression of prostate cancer by regulating MMP2 and MMP-9 [Tung MC et al, 2013].

Therefore further studies are needed to better clarify the role of NGAL in prostate cancer, as until now conflicting results have been generated between “in silico” and “in vitro” data.

### **Renal tumor**

The incidence of renal cell carcinoma (RCC) is growing [Rathmell WK et al, 2010]. Most of the patients initially diagnosed with localized disease are cured by surgery, but over 30% of them die from relapse. The molecular basis of a great diversity in clinical behavior of RCC is still unclear and makes it a target to investigate the nature of these heterogeneities [Banks RE et al, 2006].

A recent immunohistochemical study on a set of 30 surgically-resected renal tumors revealed that NGAL is expressed in several histotypes of renal tumors especially in papillary and chromphobe histotypes. NGAL expression is highest in the higher histological grade of papillary and clear cell RCC and in its peritoneal metastasis [Barresi V et al, 2010 (3)]. The authors suggested that the upregulation of NGAL in the above-mentioned tumor histotypes could be related to an increased requirement of iron uptake and could justify the use of iron chelators for renal cancer therapies. In agreement, our analysis revealed an upregulation of NGAL in RCC. Conversely, IHC evaluation did not show any immunostaining for NGAL. It was further shown that NGAL, as detected by ELISA, had predict value for progression free survival in RCC patients treated with sunitinib malate [Porta C et al, 2010]. While, most recent data conducted by Di Carlo does not reach any conclusive results on the usefulness of NGAL as a diagnostic marker [Di Carlo A, 2013].

### **Thyroid cancer**

Our data show that transcript levels of NGAL are higher in tumor thyroid cancer (papillary carcinoma) when compared with normal counterpart. In fact, 83.7% of case displayed NGAL transcripts above the 75th percentile of “normal” values, while 6.1% of cases showed mRNA levels of NGAL below the 25th percentile (Figure 5K). In contrast, no protein expression was detect by IHC (Human Protein Atlas web site). According to mRNA expression data, previous studies showed that NGAL is overexpressed in thyroid cancer [Barresi V et al, 2012 (1,2); Iannetti A et al, 2008].

## HEMATOLOGICAL MALIGNANCIES

Different than for solid tumor, NGAL expression in hematological malignancies displays a very homogeneous behavior: in fact, all the datasets that reach the levels of significance  $p < 0.01$  exhibit a uniform downregulation of NGAL compared to controls. Datasets analyzed included myeloma and leukemias (ALL, acute lymphoblastic leukemia; AML, acute myeloid leukemia, CLL; chronic lymphocytic leukemia). Accordingly, recent data show that highest mRNA NGAL levels were associated with better prognosis in AML [Yang WC et al, 2013]. According to the exclusion criteria designed for the purpose of the present study, no data on chronic myeloid leukemia (CML) were generated by computational analysis. However, NGAL was identified in an expression profiling study on CML cells. In this work, the authors analyzed gene expression profiles of cancer cells from 27 patients using a cDNA microarray. Among the 150 genes up-regulated, they observed an increase of NGAL mRNA levels in CML patients [Kaneta Y et al, 2003]. Subsequently, Villalva C et al have performed RT-PCR for NGAL expression in a large cohort of CML patients. Their results indicate that NGAL is expressed in parallel with the BCR-ABL oncoprotein at the early stage of leukemia process and it is secreted at high levels in these patients. The authors have excluded the possibility that the large increase of NGAL expression in CML patients at diagnosis resulted from the presence of circulating myelocytes in blood, which constitutively secrete NGAL protein during maturation [Villalva C et al, 2008]. Leng X et al have proposed two activities associated with NGAL in a mouse model of CML tumorigenesis. On the one hand, NGAL induces apoptosis of normal hematopoietic cells resulting in the replacement of these with leukemic cells and on the other NGAL facilitates tissue invasion through stabilization of MMP-9 activity [Leng X et al, 2008].

**Table 3. Clinical impact of NGAL expression pattern in different cancer type according to previous studies**

TUMOR TYPE	METHODS	CLINICAL IMPACT OF NGAL EXPRESSION	AUTHOR [REF.]	YEAR
<b>SOLID TUMORS</b>				
BLADDER	CM, GZ, MS	Diagnostic marker	Roy R et al	2008
			Monier F et al (1)	2000
	GZ, IHC, ELISA	Early marker of tumor progression	Monier F et al (2)	2002
BRAIN	IHC, GZ, ELISA	Diagnostic marker	Smith ER et al	2008
	IHC	Positive correlation with high proliferation index in primary tumor	Barresi V et al (4)	2010
	IHC	Associated with poor outcome	Liu MF et al	2011
BREAST	IHC	Associated with poor outcome	Stoesz SP et al	1998
	ELISA	Positive correlation with lymphatic node metastasis	Shen ZZ et al	2003
	ELISA	Positive correlation with breast cancer aggressiveness	Provatoopoulou X et al	2009
	IHC	Positive correlation with poor prognosis in primary human breast cancer	Bauer M et al	2008
	IHC	Positive correlation with poor outcome	Li SH et al	2009
	IHC	Positive correlation with poor outcome	Wenners AS et al	2010
CERVICAL	IHC	Positive correlation with HPV type	Syrjänen S et al	2012
COLORECTAL	IHC, FISH	Positive correlation with tumor transformation	Nielsen BS et al	1996
	ELISA	Prognostic utility in metastatic patients	Marti J et al (1,2)	2010,2013
	PCR, GZ	Diagnostic marker	Catalán V et al	2011
	ELISA	Not suitable as a diagnostic marker	Fung KY et al	2013
	ELISA	Not useful marker of progression	McLean MH et al	2013
ESOPHAGEAL	GZ, IHC, WB	Positive correlation to cancer differentiation	Zhang H et al	2007
	IHC	Positive correlation with progression	Du ZP et al	2011
GASTRIC	IHC, ELISA, WB	Positive correlation with poor outcome	Kubben FJ et al	2007
	IHC, ELISA	Diagnostic marker and positive correlation with poor outcome	Wang HJ et al	2010
HEAD & NECK	ELISA	Positive correlation with poor outcome	Lin CW et al	2012
HCC	IHC	Positive correlation with poor outcome	Zhang Y et al	2012
LUNG	IHC	Positive correlation with poor outcome	Friedl A et al	1999
PANCREATIC	ELISA	Diagnostic marker	Moniaux N et al	2008
OVARIAN	IHC, ELISA, WB	Promotion of epithelial to mesenchymal transition	Lim R et al	2007
	PCR, IHC, ELISA	Positive correlation to cancer differentiation	Cho H et al	2009
RENAL	IHC	Positive correlation with malignant phenotype	Barresi V et al (3)	2010
	ELISA	Positive correlation with poor outcome	Porta C et al	2010
THYROID	IHC, Real time	Positive correlation with malignant phenotype	Iannetti A et al	2008
	IHC	Diagnostic marker	Barresi V et al (1,2)	2012
<b>HEMATOLOGICAL MALIGNANCIES</b>				
AML	RT-PCR	Positive correlation with better prognosis	Yang WC et al	2013
CML	RT-PCR	Positive correlation with early stage of disease and BCR-ABL positivity	Villalva C et al	2008

**Legend:** AML, Acute Myeloid Leukemia; CM, Chromatography; CML, Chronic Myelogenous Leukemia; GZ, Gel zymography; HCC, Hepatocellular Carcinoma; IHC, Immunohistochemistry. MS, Mass Spectrometry.

## 2.2 VALIDATION OF COMPUTATIONAL RESULTS IN BLADDER CANCER

The majority of cases with TCC were by far men and aged  $\geq 65$  years (Table 4). Ever smoking was reported by 85.4% of cases and 66.4% of controls, whereas no difference was observed for education and drinking habit. Non-muscle invasive tumors (i.e. Ta/is-T1) represented 78.4% of cases, whereas papillary features was reported in 79.8% of TCCs.

**Table 4.** Distribution of 89 cases of transitional cell carcinoma of the bladder (TCC) and 119 hospital controls according to socio-demographic characteristics, tobacco smoking, alcohol drinking, and clinical pathological factors

Variables	TCCs (n=89)		Controls (n=119)		$\chi^2$ test
	n	(%)	n	(%)	
<b>Sex</b>					
Men	75	(84.3)	99	(83.2)	0.04; p=0.84
Women	14	(15.7)	20	(16.8)	
<b>Age (years)</b>					
< 65	34	(38.2)	47	(39.5)	0.15; p=0.93
65-74	39	(43.8)	49	(41.2)	
$\geq 75$	16	(18.0)	23	(19.3)	
<b>Education</b>					
< 7	42	(47.2)	53	(44.5)	0.15; p=0.93
7-11	31	(34.8)	43	(36.1)	
$\geq 12$	16	(18.0)	23	(19.3)	
<b>Smoking status</b>					
Never	13	(14.6)	40	(33.6)	27.76; p<0.01
Former	37	(41.6)	64	(53.8)	
Current	39	(43.8)	15	(12.6)	
<b>Alcohol drinking status</b>					
Never/Former	7	(7.9)	17	(14.3)	2.05; p=0.15
Current	82	(92.1)	102	(85.7)	
<b>Invasiveness<sup>a</sup></b>					
Ta/is	50	(56.8)			
T1	19	(21.6)			
T2-T4	19	(21.6)			
<b>Grading<sup>a</sup></b>					
Well/moderately differentiated	40	(45.5)			
Poorly diff./undifferentiated	48	(54.6)			
<b>Histological subtype</b>					
Papillary	71	(79.8)			
Non-papillary	18	(20.2)			

<sup>a</sup>The sum does not add up to the total because of missing values.

Urinary NGAL concentrations were significantly higher in cases than in controls (median: 18.35 vs. 7.75 ng/mg Cr; p<0.01); likewise, higher urinary levels of MMP-9 (median: 6.54 vs 1.18 ng/mg Cr; p<0.01) and MMP-9/NGAL complex (median: 0.00 vs 1.11 ng/mg Cr;

p<0.01) were observed in cases compared to controls (Table 5). Figure 7A shows increasing urinary concentrations of NGAL, MMP-9, and MMP-9/NGAL according to invasiveness, with markers concentration significantly higher in T2-T4 cases (p<0.01). Interestingly, MMP-9/NGAL complex was undetectable in 53.8% of controls, but only in 30.3% of all TCCs (5.3% of T2-T4 cases). In the univariate analysis, the expression of the three molecules was higher in poorly differentiated/undifferentiated than in well/moderately differentiated TCCs and in non-papillary than in papillary subtype (Table 5).

**Table 5.** Median urinary and serum NGAL, MMP-9, and MMP-9/NGAL complex concentrations<sup>a</sup> in 119 hospital controls and in 89 cases of transitional cell carcinoma of the bladder (TCC) according to clinical pathological features

	n	Median urinary concentrations (ng/mg Cr)			Median serum concentrations (ng/mL)		
		NGAL	MMP-9	MMP-9/NGAL complex	NGAL	MMP-9	MMP-9/NGAL complex
<b>Controls</b>	119	7.75	1.18	0.00	90.49	717.94	51.86
<b>All TCCs</b>	89	18.35	6.54	1.11	83.70	738.51	53.44
KW test		p<0.01	p<0.01	p<0.01	p=0.50	p=0.32	p=0.58
<b>Invasiveness<sup>b</sup></b>							
Ta/is	50	16.18	3.30	0.55	78.78	610.01	38.27
T1	19	15.24	5.98	0.29	84.09	822.82	57.24
T2-T4	19	68.55	24.29	6.64	139.32	943.55	113.85
KW test		p<0.01	p<0.01	p<0.01	p<0.01	p=0.01	p<0.01
Adjusted KW test		p=0.56	p=0.02	p=0.08	p=0.02	p=0.21	p=0.03
<b>Grading<sup>b</sup></b>							
Well/moderately differentiated	40	13.49	4.33	0.51	78.78	615.97	38.48
Poorly diff./undifferentiated	48	20.25	10.71	1.30	100.26	825.47	68.49
KW test		p<0.01	p<0.01	p<0.01	p=0.04	p=0.06	p=0.03
Adjusted KW test		p=0.47	p=0.04	p=0.31	p=0.76	p=0.50	p=0.71
<b>Histological subtype</b>							
Papillary	71	14.84	3.97	0.52	78.91	611.63	43.90
Non-papillary	18	49.88	44.03	10.67	113.66	937.75	75.36
KW test		p<0.01	p<0.01	p<0.01	p<0.01	p<0.01	p=0.04
Adjusted KW test		p=0.01	p=0.02	p<0.01	p=0.04	p<0.01	p=0.14

<sup>a</sup>Urinary concentrations were standardized on creatinine level (Cr) and expressed in ng/mg Cr; <sup>b</sup>The sum does not add up to the total because of missing values; <sup>c</sup>Mutually adjusted for tumour invasiveness, grading and histological subtype plus age and smoking habits. KW= Kruskal-Wallis test

However, after mutual adjustment for tumour characteristics, no association persisted with grading, whereas only MMP-9 and MMP-9/NGAL complex were still associated to tumour invasiveness.

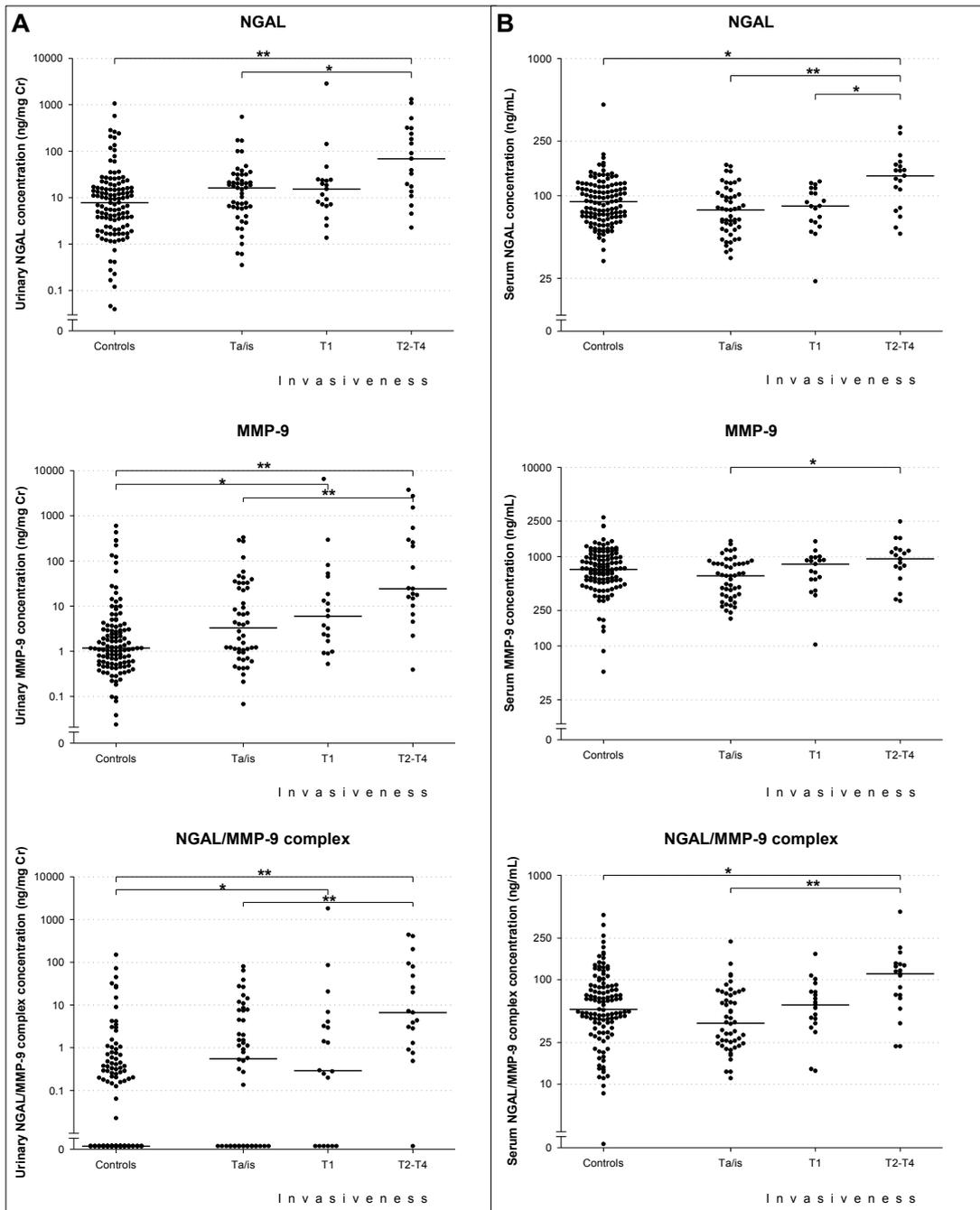
No significant differences between TCC cases and controls emerged in serum (Table 5). Nonetheless, higher levels of NGAL and MMP-9/NGAL complex were observed in patients with muscle-invasive tumors (Figure 7B). These differences were statistically different after taking into account the other tumor characteristics (Table 5).

Stronger correlations were observed between urinary and serum levels of NGAL and MMP-9 in TCC patients. These correlations were higher in patients with muscle invasive TCCs ( $r=0.72$  and  $r=0.54$ , respectively) than in those with non-muscle invasive TCCs ( $r=0.22$  and  $r=0.27$ , respectively). The correlation was less marked for MMP-9/NGAL complex. Among controls, urinary concentrations of NGAL, MMP-9, MMP-9/NGAL complex did not correlate with those in serum (Supplementary Figure 1).

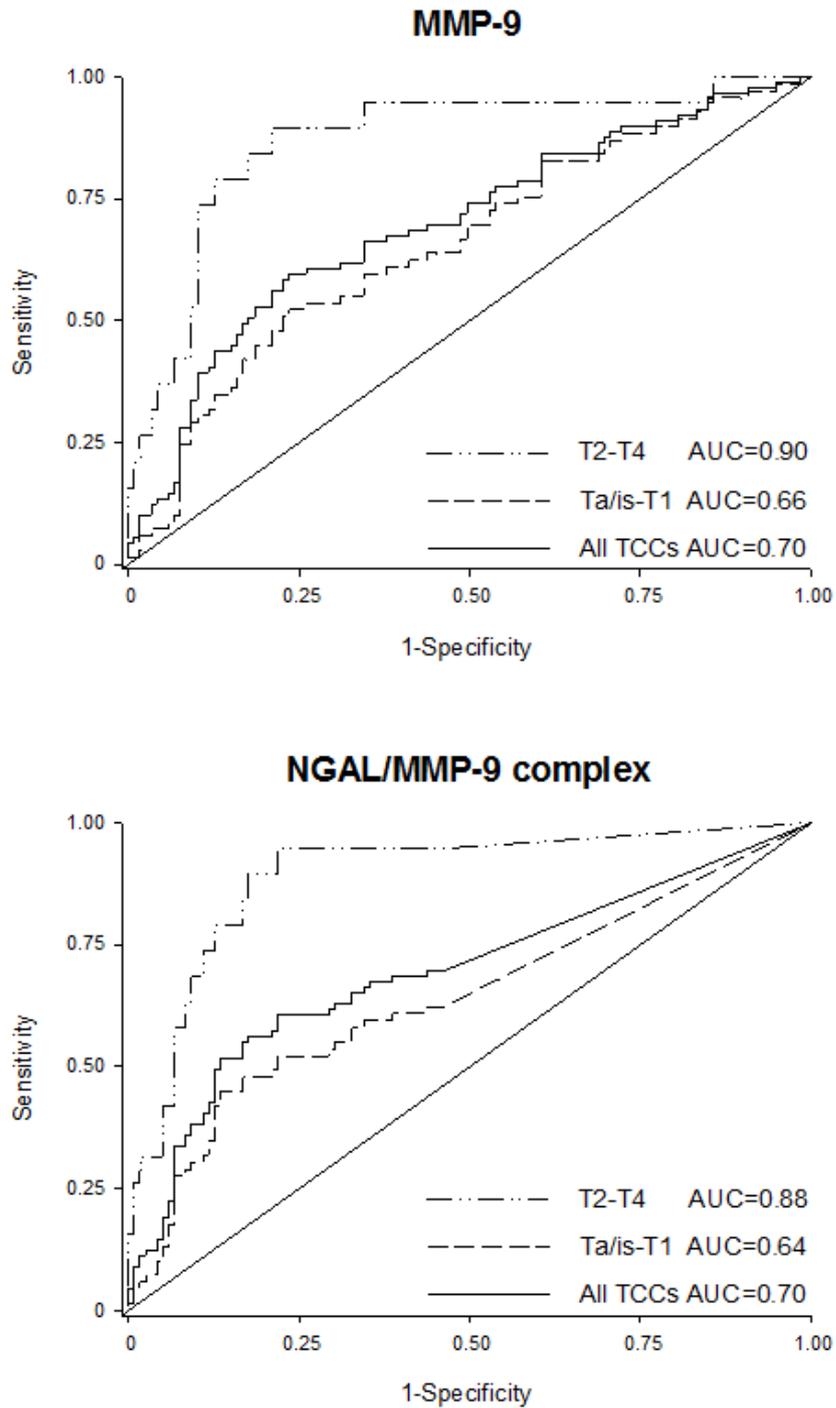
**Table 6.** Sensitivity (Se), specificity (Sp), positive predictive value (PPV), and negative predictive value (NPV) for urinary NGAL, MMP-9, and MMP-9/NGAL complex as biomarkers of transitional cell carcinoma of the bladder (TCC)

	Optimal cut-off (ng/mg Cr)	Se	Sp	PPV	NPV
<b>All TCCs</b>					
NGAL	11.59	61%	61%	53%	67%
MMP-9	2.20	65%	66%	59%	72%
MMP-9/NGAL complex	0.265	65%	67%	60%	72%
<b>Non muscle-invasive TCCs</b>					
NGAL	11.59	57%	61%	45%	71%
MMP-9	1.66	62%	58%	46%	73%
MMP-9/NGAL complex	0.19	61%	61%	48%	73%
<b>Muscle-invasive TCCs</b>					
NGAL	17.1	74%	76%	33%	95%
MMP-9	6.5	84%	82%	43%	97%
MMP-9/NGAL complex	0.89	84%	83%	44%	97%

ROC curves were used to determine the optimal cut-off for the three molecules (Table 6). Low sensitivity was reported for all the three urinary markers in all TCCs, which were able to correctly classify approximately 65% of cases. For MMP-9 and MMP-9/NGAL complex, the sensitivities and the negative predictive values greatly increased among muscle-invasive cancers (Se=84% and NPV=97%). According to ROC analysis (Figure 8), MMP-9 and MMP-9/NGAL complex were the best markers among all TCCs (AUC=0.68). For T2-T4 TCCs, MMP-9 and MMP-9/NGAL complex were still the best markers showing similar diagnostic performances (AUC=0.90 and 0.88, respectively – Table 6). Diagnostic performances in serum were generally lower than in urine (data not shown).



**Figure 7.** Distribution of NGAL, MMP-9, and MMP-9/NGAL complex concentrations in urine (A) and serum (B) in hospital controls and in cases of transitional cell carcinoma of the bladder (TCC) according to tumour invasiveness. Median values are represented by horizontal lines. P-values computed by non-parametric Kruskal-Wallis and Mann-Whitney U tests. \* indicates a p-value < 0.05; \*\* indicates a p-value < 0.01.



**Figure 8.** Receiver operating characteristic (ROC) curves for urinary MMP-9 and MMP-9/NGAL complex concentrations, according to tumor invasiveness.

### 3. DISCUSSION

In the present study, computational and IHC evaluations were performed to understand whether differences in NGAL transcript or protein levels occur in different cancer types when compared with the relative normal tissues or the metastatic counterparts.

Our computational analysis suggest an active role of NGAL in early stages of tumor development, reason that many authors proposed NGAL as a diagnostic and prognostic marker. While a decrease of NGAL expression in metastatic samples was detected when compared to matched primary tumors. This observation suggests a common inactivation pathway of NGAL gene during distant tumor dissemination and leads us to venture the hypothesis that NGAL could play a protective role in metastatic development.

In particular, the tumors showing higher NGAL expression (expression levels greater than the 75th percentile of “normal” samples) were: ovarian (91.2%), thyroid (83.7%), liver (68.8%), colon (66.3%), kidney (64.7%), lung (63.1%), pancreas (60.2%) and bladder (50.5%). While, the percentages of tumor cases showing NGAL transcripts below the 25th percentile of “normal” values were almost 100% of all hematological malignancies and 94.6% of head and neck cancer, 71.4% of esophagus cancer and 59.4% of cervical carcinoma (Figure 5). These data suggest that NGAL is a candidate marker for tumor growth in a fraction of solid tumor and a favorable prognostic factor for the remaining cancer types showing lower levels of NGAL transcript levels.

Validation of computational studies, above described, was performed on bladder cancer. In developed countries, transitional cell carcinoma of the bladder (TCC) is the fourth most frequent cancer in men, with the highest incidence worldwide in southern Europe [Jemal A et al, 2011]. The major risk factors for bladder cancer are cigarette smoking and occupational exposure to aromatic amines; while, inconclusive epidemiological data have been generated on diet. On the one hand, dietary intake of vegetables and/or fruits protect against bladder cancer, on the other hand genetic variability may alter such association [Tang L, et al 2010; Lin J, et al, 2009].

Several markers have been proposed in recent years, but none is currently considered adequate to diagnose and predict the outcome of bladder cancer. The identification of novel factors, such as NGAL, might be useful in the management of bladder cancer and might reduce invasive procedures, such as cystoscopy that remains expensive, painful, and cause

distress for patients [Almallah YZ et al 2000, Yerlikaya G et al, 2014] reducing the compliance with follow-up.

These validating data showed an association between NGAL, MMP-9 and MMP-9/NGAL complex and TCC, and the associations were consistent with respect to possible perturbation due to lifestyle factors. As expected, these three biomarkers showed higher diagnostic properties in urine than in serum; indeed, TCC is localized in the inner layer the bladder where it can excrete these proteins directly in the urine. These findings are also supported by our previous observation in which the immunostaining of NGAL reveals its localization in bladder cancer cells [Candido S et al, 2014].

Multiple proteins have been measured in bladder cancer patients showing the specificity of the analysis in biological fluid such as urine. Among these proteins, in agreement with our findings, the authors revealed a strong association of higher MMPs urine levels with invasiveness and grading [Rosser CJ et al, 2013; Eissa S et al 2007; Gerhards S et al , 2001; Nutt JE et al, 2003; Fernández CA et al, 2009; Sier CFM et al, 2000]. A recent study on renal cell carcinomas compared NGAL, MMP-9 and MMP-9/NGAL complex in urine and serum [Di Carlo A, 2013]. Expression levels of NGAL were strongly correlated in both urine and serum from these patients. However, the authors failed to demonstrate such correlation for MMP-9 and MMP-9/NGAL complex levels. Conversely, the present study showed a strong correlation between serum and urine levels of NGAL, MMP-9, and MMP-9/NGAL complex only in TCCs with an aggressive phenotype showing their role in invasiveness (Supplementary Figure 1). Accordingly, MMP-9 and MMP-9/NGAL complex showed sensitivity and specificity higher than 80% for muscle-invasive TCCs. NPV was particularly elevated, that is the probability of having the disease given a negative test is very low (3% for MMP-9 and 4% for MMP-9/NGAL complex). On the other hand, positive predictive values were between 55% and 73%, suggesting a moderate capacity to identify cases. These results suggested that these molecules could be used as exclusion test. Several investigations have previously reported similar results for MMP-9 in bladder cancer [Rosser CJ et al, 2013; Eissa S et al 2007; Gerhards S et al , 2001; Nutt JE et al, 2003; Fernández CA et al, 2009; Sier CFM et al, 2000]. However, these studies were heterogeneous according to tumor characteristics (e.g., histological type, stage, grade), and none of them have reported the prognostic properties according to tumor stage. The faculty to detect a disease at an early stage is a particularly interesting aspect in the evaluation of a new biomarker. Indeed, we found poor sensitivity and specificity for non-muscle invasive TCCs, suggesting that the

overall diagnostic properties of the three molecules were driven by T2-T4 cancers. Similar results were reported by Gerhards and colleagues for urinary MMP-2 [Gerhards S et al ,2001].

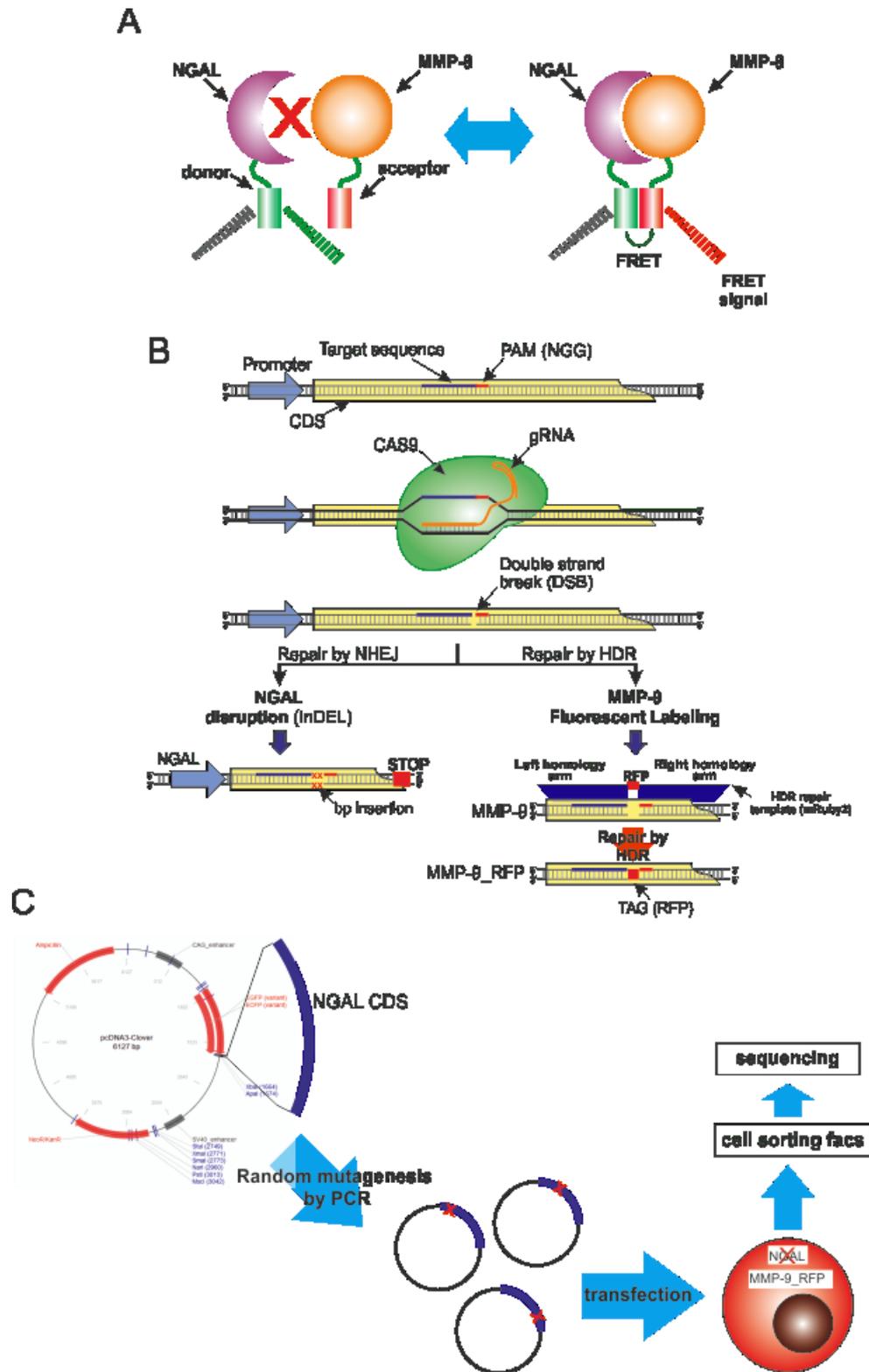
An efficient tumour biomarker is expected to be cost-effective in the detection of cancer at an early stage, and in the discrimination between low-risk and high-risk cancers [Larré S et al, 2013]. According to this point of view, our finding may help to better define the diagnostic properties of NGAL, MMP-9, and MMP-9/NGAL complex. Firstly, the correlation was stronger in urine rather than in serum, suggesting that urine is the most adequate among body fluids for these biomarkers. Secondly, the three proteins correlated with tumour invasiveness, being therefore able to discriminate low-risk from high-risk cancers. However, sensitivities and positive predictive values were very low in non-muscle invasive TCCs, bringing to light the limits of these markers to detect early-stage cancers.

A strength of this study was the use of hospital-controls, since, following a pragmatic approach, it could give more reliable information on the actual diagnostic properties of these biomarkers. Further, the availability of information on several lifestyle factors, including diet habits, was an additional strength. Indeed, possible alteration of markers concentration due to lifestyle factors was evaluated and markers performances were tested taking into account this possible source of bias. Several previous investigation considered volunteers or healthy people as control group [Eissa S et al, 2007; Sier CFM et al, 2000], but this choice is prone to selection bias [Behrens T et al, 2014] and may artificially increase the markers specificity. Indeed, volunteers are known to be generally healthier than the general population, thus reducing the number of false positives. Moreover, other advantages derive from the case control-study design [Polesel J et, 2012]. First, the use of matched controls may have prevented differences in protein concentration due to dissimilarities between cases and controls in relation to age and/or gender (i.e., matching characteristics). Then, patients were approached during their hospital staying, limiting selection bias and ensuring that urine and blood samples were collected by trained nurses prior to any cancer treatment, adhering to standard clean-catch procedure.

In conclusion, the present study deepens the knowledge of the molecular mechanisms sustaining NGAL expression in tumor cells and its effects on cancer metastatic behavior. Furthermore, our experimental results suggested that MMP-9/NGAL pathway is associated with an aggressive phenotype of TCC, although further confirmations are needed. These findings suggest that these proteins may be integrated in the surveillance of bladder cancer, thus improving patients complaints and diminishing their discomfort.

### **3.4 FUTURE CHALLENGES**

In vitro and in vivo studies are needed to better clarify the mechanism of interaction of MMP-9 and NGAL and to lay the groundwork for the development of new drugs that could inhibit their interaction. In the Figure 9 is clearly described which molecular approaches will be used for a deep understanding of such interaction. NGAL eGFP mutant library (Figure 9C) will be used to infect a modified bladder cancer cell line null for NGAL and carrying mRuby2 reporter in the MMP-9 gene/protein, obtained by Crispr/Cas technology [Choi PS et al, 2014] (Figure 9B). Förster Resonance Energy Transfer (FRET) technology [Lam AJ et al, 2012] will be used to isolate clones that not show FRET signal because of the failure of interaction between MMP-9-mRuby2 and NGAL-eGFP proteins (Figure 9A). The selected mutations of NGAL, identified by sequencing assay, will be introduced in bladder cell models by Crispr/Cas [Choi PS et al, 2014] to confirm in vitro and in vivo the failure of MMP-9/NGAL interaction and to evaluate the resulting change on gelatinolytic activity of MMP-9.



**Figure 9. (A)** Identification of Mutant Clones of NGAL that Lose the Ability to Bind MMP-9 using FRET assay to isolate clones that not show FRET signal because of the failure of interaction between MMP-9-mRuby2 and NGAL-eGFP proteins **(B)** NGAL Knockdown and Fluorescent Labeling of MMP-9 gene obtained using Crispr/Cas Technology. **(C)** The NGAL-null cells carrying mRuby2 reporter are infected with the plasmid pool of NGAL Clover mutant library, obtained using random mutagenesis by PCR. Cell sorting is performed to select cell clones that not show FRET signal. Sequencing of exogenous NGAL insects of each selected cell clone is performed to identify the NGAL point mutations, including nonsense mutations, which may inhibit the formation of MMP-9/NGAL complex.

## 4. MATERIAL AND METHODS

### 4.1 COMPUTATIONAL ANALYSES

ONCOMINE software (<https://www.oncomine.com>) was used (December 2013) to compare mRNA expression levels between normal tissue versus tumor and primary tumor versus metastatic, and in both cases evaluated on biopsy samples. Statistical analysis of the differences in NGAL mRNA expression between the abovementioned sets of samples was accomplished through use of ONCOMINE algorithms. Only datasets generated by Affymetrix U133 platform were considered for the present analysis. Of note, this platform used the “212531\_at” NGAL probe set. Datasets showing different expression analysis between normal and tumor tissues with statistical significance less than 0.01 (by t-test) and fold change  $\leq -2$  or  $\geq 2$  were included; while, those showing a differential expression between primary tumor and metastasis with a statistic significance less than 0.05 (by t-test) and fold change  $\leq -1.5$  or  $\geq 1.5$  were considered. According to these criteria, 38 datasets were used for the purpose of the study. Of these, 29 were used for the comparative analysis between normal and tumor tissues (Table 1); while, 9 were used to analyze the differences between primary tumor and metastasis (Table 2). Fold change was calculated to evaluate the changes in gene expression between the groups of samples included in this analysis. Properties of the datasets are presented in Tables 1 and 2.

To assess the distribution of NGAL transcript levels among cancer types and normal samples, mRNA expression levels from Oncomine analysis were normalized using an housekeeping gene in each dataset and then merged together. The choice of the housekeeping gene was made on the basis of the homogenous mRNA levels distribution. When more than one probset was available, the mean value of the same housekeeping gene was used for the normalization. To define the “normal range” of expression we calculated the 25th and the 75th percentile of NGAL transcript levels in normal samples for each tumor type. Accordingly, the percentage of tumor samples showing NGAL levels outside the defined “normal range” was calculated.

NGAL protein expression was performed by analyzing the web site of Human Protein Atlas (<http://www.proteinatlas.org/>). As indicate in the web site, expression of NGAL for each tumor type was evaluated as strong, moderate, weak and negative immunostaining. The Sigma-Aldrich HPA002695 NGAL antibody was employed for this analysis.

## **4.2 PATIENTS RECRUITMENT AND BLOOD SAMPLE COLLECTION**

The present data derived from a case-control study was conducted from 2004 to 2009 on TCCs within an established Italian network of collaborating centres. This analysis included the first cases enrolled up to August 2007 in the province of Pordenone whom urine and blood samples were available [Polesel J et al, 2012]. Cases were 89 patients aged 18 years or older (median age: 66 year) with incident histologically or cytologically confirmed TCC admitted to major general hospitals. Using the TNM classification, cases were classified in non-muscle invasive (i.e., Ta/is-T1) and muscle invasive (i.e., T2-T4) tumors according to the guidelines of European Association of Urology [Babjuk M et al, 2013].

The control group included 119 patients (median age: 66 year) admitted to the same network of hospitals for a wide spectrum of acute, non-neoplastic conditions unrelated to tobacco and alcohol consumption, to known risk factors for the bladder cancer, or to conditions associated to long-term diet modification. All study subjects signed an informed consent, according to the recommendations of the Board of Ethics of the study hospitals.

Trained nurses administered a validated, structured questionnaire [D'Avanzo B et al, 1996] to cases and controls during their hospital stay, thus keeping refusal below 5% for both cases and controls. The questionnaire collected information on socio-demographic factors and life-style factors, including smoking and alcohol drinking habits. Each patient enrolled in the study provided peripheral blood and urine samples on the day they were interviewed. Samples were collected before patients had undergone any treatment. Standard clean-catch procedure for urine collection (50-mL sample of first-voided for each patients) was performed to prevent sample contamination. Half of the sample (25 mL) was immediately frozen at -80°C and the remaining one was stored in CytoLyt solution at 4°C. Blood samples were centrifuged at 1,500g for 10 minutes obtaining serum, buffy coat, and red blood cells and then stored at -80°C. Serum, plasma and urine samples were stored at -80°C until analyses.

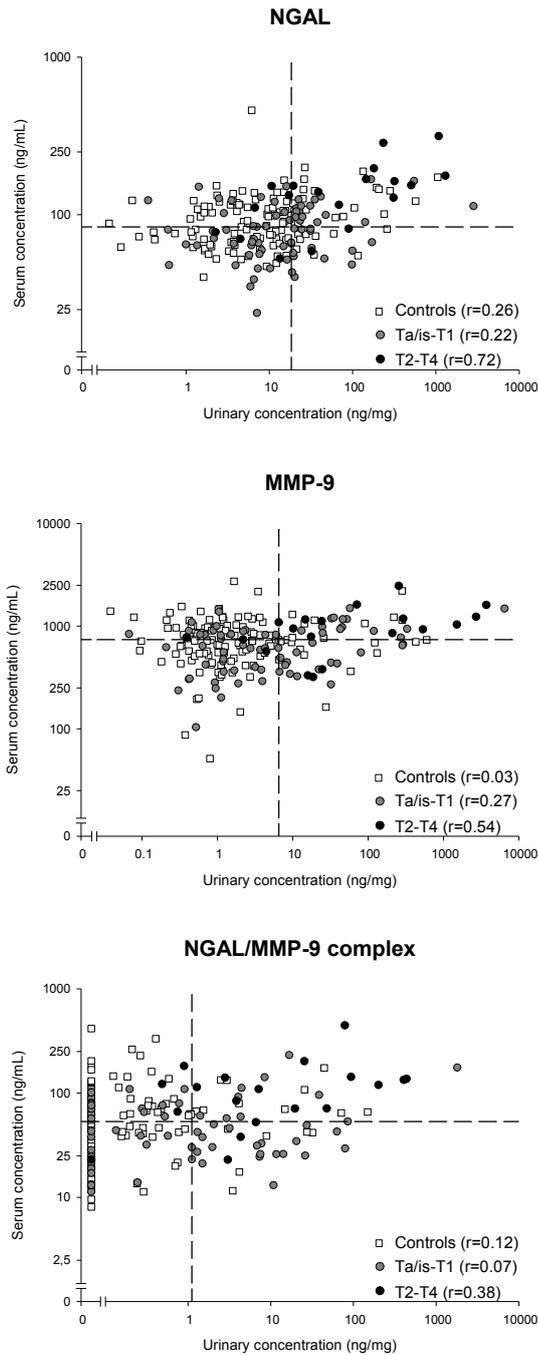
## **4.3 ELISA ASSAY**

Serum and urine concentration of MMP-9, NGAL and MMP-9/NGAL complex were assayed, according to manufacturers' protocols, by specific, commercially available, through enzyme-linked immunosorbent (ELISA) assay kits (Quantikine, R&D Systems Inc.,USA) in accordance with the manufacturer's instructions and analyzed with an ELISA reader (Tecan Systems) at 450 nm. Urinary concentrations were standardized according to creatinine level

(Cr) and expressed as ng/mg Cr. Urinary Creatinine was assayed using ABX Pentra Enzymatic Creatinine CP kit (HORIBA ABX INC, USA) according to manufacturer's instructions. Colorimetric intensity was assayed at 545 nm using ABX Pentra 400 analyzer (HORIBA ABX INC, USA). All analyses were carried out at the Department of Biomedical and Biotechnological Sciences, University of Catania.

#### **4.4 STATISTIC ANALYSIS**

Differences of urinary and serum levels of NGAL, MMP-9, and MMP-9/NGAL complex according to tumor characteristics were evaluated using the nonparametric Kruskal-Wallis test, followed by Dunn's multiple comparison post-test. To evaluate the independent effect of each tumor feature, a multivariate Kruskal-Wallis test was further adopted [May WL et al, 1997]. For each molecule, the agreement between urinary and serum concentrations was measured through Spearman's rank correlation coefficient. The performance of these proteins as cancer biomarkers was evaluated in terms of sensitivity and specificity, overall and according to tumor characteristics. Receiver operating characteristic (ROC) analysis was performed to determine the optimal cut-off for diagnostic purpose and sensitivity (Se), specificity (Sp), positive predictive value (PPV), and negative predictive value (NPV) were calculated. Discrimination was quantified by the area under the ROC curve (AUC) [Faraggi D et al, 2002].



**Supplementary Figure 1.** Correlation between urinary and serum concentrations of NGAL, MMP-9, and MMP-9/NGAL complex in hospital controls and in cases of transitional cell carcinoma of the bladder (TCC). Median values for TCCs are represented by short-dotted lines

## 5. REFERENCES

- Agnelli L, Mosca L, Fabris S, Lionetti M, Andronache A, Kwee I, Todoerti K, Verdelli D, Battaglia C, Bertoni F, Delilieri GL, Neri A. A SNP microarray and FISH-based procedure to detect allelic imbalances in multiple myeloma: an integrated genomics approach reveals a wide gene dosage effect. *Genes Chromosomes Cancer*. 2009;48:603–14.
- Ahmedin J, Thomas A, Murray T, Thun M. Cancer statistics, 2002. *CA Cancer J Clin*. 2002;52:23–42.
- Almallah YZ, Rennie CD, Stone J, Lancashire MJR. Urinary tract infection and patient satisfaction after flexible cystoscopy and urodynamic evaluation. *Urology* 2000;56:37-39.
- Anglesio MS, Arnold JM, George J, Tinker AV, Tothill R, Waddell N, Simms L, Locandro B, Fereday S, Traficante N, Russell P, Sharma R, Birrer MJ, AOCs Study Group, deFazio A, Chenevix-Trench G, Bowtell DD. Mutation of ERBB2 provides a novel alternative mechanism for the ubiquitous activation of RAS-MAPK in ovarian serous low malignant potential tumors. *Mol Cancer Res*. 2008;6:1678–90.
- Argani P, Rosty C, Reiter RE, Wilentz RE, Murugesan SR, Leach SD, Ryu B, Skinner HG, Goggins M, Jaffee EM, Yeo CJ, Cameron JL, Kern SE, et al. Discovery of new markers of cancer through serial analysis of gene expression: prostate stem cell antigen is overexpressed in pancreatic adenocarcinoma. *Cancer Res*. 2001;61:4320–4324.
- Babjuk M, Burger M, Zigeuner R, Shariat SF, van Rhijn BWG, Compérat E, Sylvester RJ, Kaasinen E, Böhle A, Redorta JP, Rouprêt. EAU guidelines on non-muscle invasive urothelial carcinoma of the bladder: Update 2013. *Eur Urol* 2013; 64:639-653.
- Badea L, Herlea V, Dima SO, Dumitrascu T, Popescu I. Combined gene expression analysis of whole-tissue and microdissected pancreatic ductal adenocarcinoma identifies genes specifically overexpressed in tumor epithelia. *Hepatogastroenterology*. 2008;55:2016–27.
- Banks RE, Tirukonda P, Taylor C, Hornigold N, Astuti D, Cohen D, Maher ER, Stanley AJ, Harnden P, Joyce A, Knowles M, Selby PJ. Genetic and epigenetic analysis of von Hippel-Lindau (VHL) gene alterations and relationship with clinical variables in sporadic renal cancer. *Cancer Res*. 2006;66:2000–11.

- Barresi V (1), Vitarelli E, Reggiani Bonetti L, Tuccari G, Barresi G. Diagnostic value of neutrophil gelatinase-associated lipocalin (NGAL) immunoexpression in follicular-patterned lesions of the thyroid gland. *Virchows Arch.* 2012;460:319–25.
- Barresi V (2), Leni A, Tuccari G, Barresi G. Neutrophil gelatinase-associated lipocalin (NGAL) immunohistochemical expression in follicular cell-derived thyroid tumors: a novel diagnostic tool? *Histol Histopathol.* 2012;27:329–36.
- Barresi V (3), Ieni A, Bolignano D, Magno C, Buemi M, Barresi G. Neutrophil gelatinase-associated lipocalin immunoexpression in renal tumors: correlation with histotype and histological grade. *Oncol Rep.* 2010;24:305–10.
- Barresi V (4), Tuccari G, Barresi G. NGAL immunohistochemical expression in brain primary and metastatic tumors. *Clin Neuropathol.* 2010;29:317–22.
- Bauer M, Eickhoff JC, Gould MN, Mundhenke C, Maass N, Friedl A. Neutrophil gelatinase-associated lipocalin (NGAL) is a predictor of poor prognosis in human primary breast cancer. *Breast Cancer Res Treat.* 2008;108:389–97.
- Behrens T, Bonberg N, Casjens S, Pesch B, Brüning T. A practical guide to epidemiological practice and standards in the identification and validation of diagnostic markers using a bladder cancer example. *Biochim Biophys Acta* 2014;1844:145-155.
- Berger T, Cheung CC, Elia AJ, Mak TW. Disruption of the *Lcn2* gene in mice suppresses primary mammary tumor formation but does not decrease lung metastasis. *Proc Natl Acad Sci U S A.* 2010;107:2995–3000.
- Bolignano D, Donato V, Lacquaniti A, Fazio MR, Bono C, Coppolino G, Buemi M. Neutrophil gelatinase-associated lipocalin (NGAL) in human neoplasias: a new protein enters the scene. *Cancer Lett.* 2010;288:10–6.
- Bonome T, Levine DA, Shih J, Randonovich M, Pise-Masison CA, Bogomolny F, Ozbun L, Brady J, Barrett JC, Boyd J, Birrer MJ. A gene signature predicting for survival in suboptimally debulked patients with ovarian cancer. *Cancer Res.* 2008;68:5478–86.

- Bousserouel S, Kauntz H, Gossé F, Bouhadjar M, Soler L, Marescaux J, Raul F. Identification of gene expression profiles correlated to tumor progression in a preclinical model of colon carcinogenesis. *Int J Oncol.* 2010;36:1485–90.
- Braun V, Braun M. Active transport of iron and siderophore antibiotics. *Curr Opin Microbiol.* 2002;5:194–201.
- Candido S, Abrams SL, Steelman LS, Lertpiriyapong K, Fitzgerald TL, Martelli AM, Cocco L, Montalto G, Cervello M, Polesel J, Libra M, McCubrey JA. Roles of NGAL and MMP-9 in the tumor microenvironment and sensitivity to targeted therapy. *Biochim Biophys Acta* 2015. [Epub ahead of print]
- Candido S, Maestro R, Polesel J, Catania A, Maira F, Signorelli SS, McCubrey JA, Libra M. Roles of neutrophil gelatinase-associated lipocalin (NGAL) in human cancer. *Oncotarget* 2014;30:1576-94.
- Catalán V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Silva C, Rotellar F, Hernández-Lizoain JL, Baixauli J, Valentí V, Pardo F, Salvador J, Frühbeck G. Up-regulation of the novel proinflammatory adipokines lipocalin-2, chitinase-3 like-1 and osteopontin as well as angiogenic-related factors in visceral adipose tissue of patients with colon cancer. *J Nutr Biochem.* 2011;22:634–41.
- Chakraborty S, Kaur S, Guha S, Batra SK. The multifaceted roles of neutrophil gelatinase associated lipocalin (NGAL) in inflammation and cancer. *Biochim Biophys Acta.* 2012;1826:129–69.
- Chappell WH (1), Abrams SL, Montalto G, Cervello M, Martelli AM, Candido S, Libra M, Polesel J, Talamini R, Arlinghaus R, Steelman LS, McCubrey JA. Effects of ectopic expression of NGAL on doxorubicin sensitivity. *Oncotarget.* 2012;3:1236–45.
- Chappell WH (2), Abrams SL, Franklin RA, LaHair MM, Montalto G, Cervello M, Martelli AM, Nicoletti F, Candido S, Libra M, Polesel J, Talamini R, Milella M, et al. Ectopic NGAL expression can alter sensitivity of breast cancer cells to EGFR, Bcl-2, CaM-K inhibitors and the plant natural product berberine. *Cell Cycle.* 2012;11:4447–61.

- Chen YJ, Chang LS. NF $\kappa$ B- and AP-1-mediated DNA looping regulates matrix metalloproteinase-9 transcription in TNF- $\alpha$ -treated human leukemia U937 cells. *Biochim Biophys Acta*. 2015 Oct;1849(10):1248-1259.
- Cho H, Kim JH. Lipocalin 2 expressions correlate significantly with tumor differentiation in epithelial ovarian cancer. *J Histochem Cytochem*. 2009;57:513–21.
- Choi PS, Meyerson M. Targeted genomic rearrangements using CRISPR/Cas technology. *Nat Commun*. 2014; 5: 3728.
- Cooperman AM. Pancreatic cancer: the bigger picture. *Surg Clin North Am*. 2001;81:557–74.
- Cowland JB, Muta T, Borregaard N. IL-1beta-specific up-regulation of neutrophil gelatinase-associated lipocalin is controlled by IkappaB-zeta. *J Immunol*. 2006;176:5559–66.
- Cowland JB, Sørensen OE, Sehested M, Borregaard N. Neutrophil gelatinase-associated lipocalin is up-regulated in human epithelial cells by IL-1 beta, but not by TNF-alpha. *J Immunol*. 2003;171:6630–9.
- D'Avanzo B, La Vecchia C, Katsouyanni K, Negri E, Trichopoulos D. Reliability of information on cigarette smoking and beverage consumption provided by hospital controls. *Epidemiology* 1996;7:312–315.
- Devireddy LR, Gazin C, Zhu X, Green MR. A cell-surface receptor for lipocalin 24p3 selectively mediates apoptosis and iron uptake. *Cell*. 2005;123:1293–305.
- Di Carlo A. Evaluation of neutrophil gelatinase-associated lipocalin (NGAL), matrix metalloproteinase-9 (MMP-9) and their complex MMP-9/NGAL in sera and urine of patients with kidney tumours. *Oncol Lett* 2013;5:1677-81.
- Du ZP, Lv Z, Wu BL, Wu ZY, Shen JH, Wu JY, Xu XE, Huang Q, Shen J, Chen HB, Li EM, Xu LY. Neutrophil gelatinase-associated lipocalin and its receptor: independent prognostic factors of oesophageal squamous cell carcinoma. *J Clin Pathol*. 2011;64:69–74.
- Eissa S, Ali-Labib R, Swellam M, Bassiony M, Tash F, El-Zayat TM. Noninvasive diagnosis of bladder cancer by detection of matrix metalloproteinases (MPP-2 and MMP-9) and their inhibitor (TIMP-2) in urine. *Eur Urology* 2007;52:1388-97.

- Fang WK, Xu LY, Lu XF, Liao LD, Cai WJ, Shen ZY, Li EM. A novel alternative spliced variant of neutrophil gelatinase-associated lipocalin receptor in oesophageal carcinoma cells. *Biochem J.* 2007;403:297–303.
- Faraggi D, Reiser B. Estimation of the area under the ROC curve. *Stat Med* 2002; 21:3093-3016.
- Fernández CA, Eszolek MF, Loughlin KR, Libertino JA, Summerhayes IC, Shuber AP. A novel approach to using matrix metalloproteinases for bladder cancer. *J Urol* 2009; 182:2188-2194.
- Fernández CA, Yan L, Louis G, Yang J, Kutok JL, Moses MA. The matrix metalloproteinase-9/neutrophil gelatinase-associated lipocalin complex plays a role in breast tumor growth and is present in the urine of breast cancer patients. *Clin Cancer Res.* 2005;11:5390–5.
- Flower DR. The lipocalin protein family: structure and function. *Biochem J.* 1996;318:1–14.
- Friedl A, Stoesz SP, Buckley P, Gould MN. Neutrophil gelatinase-associated lipocalin in normal and neoplastic human tissues. Cell type-specific pattern of expression. *Histochem J.* 1999;31:433–41.
- Fung KY, Priebe I, Purins L, Tabor B, Brierley GV, Lockett T, Nice E, Gibbs P, Tie J, McMurrick P, Moore J, Ruszkiewicz A, Burgess A, Cosgrove LJ. Performance of serum lipocalin 2 as a diagnostic marker for colorectal cancer. *Cancer Biomark.* 2013;13:75–9.
- Gerhards S, Jung K, Koenig F, Daniltchenko D, Hauptmann S, Schnorr D, Loening SA. Excretion of matrix metalloproteinases 2 and 9 in urine is associated with high stage and grade of bladder carcinoma. *Urology* 2001;57:675-79.
- Giordano TJ, Au AY, Kuick R, Thomas DG, Rhodes DR, Wilhelm KG, Jr, Vinco M, Misek DE, Sanders D, Zhu Z, Ciampi R, Hanash S, Chinnaiyan A, et al. Delineation, functional validation, and bioinformatic evaluation of gene expression in thyroid follicular carcinomas with the PAX8-PPARG translocation. *Clin Cancer Res.* 2006;12:1983–93.
- Goetz DH, Holmes MA, Borregaard N, Bluhm ME, Raymond KN, Strong RK. The neutrophil lipocalin NGAL is a bacteriostatic agent that interferes with siderophore-mediated iron acquisition. *Mol Cell*- 2002;10:1033–43.

- Haferlach T, Kohlmann A, Wieczorek L, Basso G, Kronnie GT, Béné MC, De Vos J, Hernández JM, Hofmann WK, Mills KI, Gilkes A, Chiaretti S, Shurtleff SA, et al. Clinical utility of microarray-based gene expression profiling in the diagnosis and subclassification of leukemia: report from the International Microarray Innovations in Leukemia Study Group. *J Clin Oncol*. 2010;28:2529–37.
- Han H, Bearss DJ, Browne LW, Calaluce R, Nagle RB, Von Hoff DD. Identification of differentially expressed genes in pancreatic cancer cells using cDNA microarray. *Cancer Res*. 2002;62:2890–2896.
- He H, Jazdzewski K, Li W, Liyanarachchi S, Nagy R, Volinia S, Calin GA, Liu CG, Franssila K, Suster S, Kloos RT, Croce CM, de la Chapelle A. The role of microRNA genes in papillary thyroid carcinoma. *Proc Natl Acad Sci U S A*. 2005;102:19075–80.
- Hendrix ND, Wu R, Kuick R, Schwartz DR, Fearon ER, Cho KR. Fibroblast growth factor 9 has oncogenic activity and is a downstream target of Wnt signaling in ovarian endometrioid adenocarcinomas. *Cancer Res*. 2006;66:1354–62.
- Holmes MA, Paulsene W, Jide X, Ratledge C, Strong RK. Siderocalin (Lcn 2) also binds carboxymycobactins, potentially defending against mycobacterial infections through iron sequestration. *Structure*. 2005;13:29–41.
- Hong Y, Downey T, Eu KW, Koh PK, Cheah PY. A ‘metastasis-prone’ signature for early-stage mismatch-repair proficient sporadic colorectal cancer patients and its implications for possible therapeutics. *Clin Exp Metastasis*. 2010;27:83–90.
- Hu L, Hittelman W, Lu T, Ji P, Arlinghaus R, Shmulevich I, Hamilton SR, Zhang W. NGAL decreases E-cadherin-mediated cell-cell adhesion and increases cell motility and invasion through Rac1 in colon carcinoma cells. *Lab Invest*. 2009;89:531–48.
- Hu N, Clifford RJ, Yang HH, Wang C, Goldstein AM, Ding T, Taylor PR, Lee MP. Genome wide analysis of DNA copy number neutral loss of heterozygosity (CNNLOH) and its relation to gene expression in esophageal squamous cell carcinoma. *BMC Genomics*. 2010;11:576.
- Huang JQ, Zheng GF, Sumanac K, Irvine EJ, Hunt RH. Meta-analysis of the relationship between cagA seropositivity and gastric cancer. *Gastroenterology*. 2003;125:1636–1644.

- Iannetti A, Pacifico F, Acquaviva R, Lavorgna A, Crescenzi E, Vascotto C, Tell G, Salzano AM, Scaloni A, Vuttariello E, Chiappetta G, Formisano S, Leonardi A. The neutrophil gelatinase-associated lipocalin (NGAL), a NF-kappaB-regulated gene, is a survival factor for thyroid neoplastic cells. *Proc Natl Acad Sci U S A*. 2008;105:14058–63.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global Cancer Statistics. *Ca Cancer J Clin* 2011;61:69-90.
- Jones J, Otu H, Spentzos D, Kolia S, Inan M, Beecken WD, Fellbaum C, Gu X, Joseph M, Pantuck AJ, Jonas D, Libermann TA. Gene signatures of progression and metastasis in renal cell cancer. *Clin Cancer Res*. 2005;11:5730–9.
- Kaiser S, Park YK, Franklin JL, Halberg RB, Yu M, Jessen WJ, Freudenberg J, Chen X, Haigis K, Jegga AG, Kong S, Sakthivel B, Xu H, et al. Transcriptional recapitulation and subversion of embryonic colon development by mouse colon tumor models and human colon cancer. *Genome Biol*. 2007;8:R131.
- Kaneta Y, Kagami Y, Tsunoda T, Ohno R, Nakamura Y, Katagiri T. Genome-wide analysis of gene-expression profiles in chronic myeloid leukemia cells using a cDNA microarray. *Int J Oncol*. 2003;23:681–691.
- Karlsen JR, Borregaard N, Cowland JB. Induction of neutrophil gelatinase-associated lipocalin expression by co-stimulation with interleukin-17 and tumor necrosis factor-alpha is controlled by IkappaB-zeta but neither by C/EBP-beta nor C/EBP-delta. *J Biol Chem*. 2010;285:14088–100.
- Kjeldsen L, Johnsen A, Sengeløv H, Borregaard N. Isolation and primary structure of NGAL, a novel protein associated with human neutrophil gelatinase. *J Biol Chem*. 1993;268:10425–10432.
- Koshy M, Esiashvilli N, Landry JC, Thomas CR, Jr, Matthews RH. Multiple management modalities in esophageal cancer: epidemiology, presentation and progression, work-up, and surgical approaches. *Oncologist*. 2004;9:137–46.
- Krewulak KD, Vogel HJ. Structural biology of bacterial iron uptake. *Biochim Biophys Acta*. 2008;1778:1781–1804.

- Kubben FJ, Sier CF, Hawinkels LJ, Tschesche H, Van Duijn W, Zuidwijk K, van der Reijden JJ, Hanemaaijer R, Griffioen G, Lamers CB, Verspaget HW. Clinical evidence for a protective role of lipocalin-2 against MMP-9 autodegradation and the impact for gastric cancer. *Eur J Cancer*. 2007;43:1869–76.
- Kulis M, Queirós AC, Beekman R, Martín-Subero JI. Intragenic DNA methylation in transcriptional regulation, normal differentiation and cancer. *Biochim Biophys Acta*. 2013; 1829:1161-74.
- Lam AJ, St-Pierre F, Gong Y, Marshall JD, Cranfill PJ, Baird MA, McKeown MR, Wiedenmann J, Davidson MW, Schnitzer MJ, Tsien RY, Lin MZ. Improving FRET dynamic range with bright green and red fluorescent proteins. *Nat Methods*. 2012; 9:1005-12.
- Landi MT, Dracheva T, Rotunno M, Figueroa JD, Liu H, Dasgupta A, Mann FE, Fukuoka J, Hames M, Bergen AW, Murphy SE, Yang P, Pesatori AC, et al. Gene expression signature of cigarette smoking and its role in lung adenocarcinoma development and survival. *PLoS*. 2008;3:e1651.
- Larré S, Catto JWF, Cookson MS, Messing EM, Shariat SF, Soloway MS, Svatek RS, Lotan Y, Zlotta AR, Grossman HB. *Eur Urol* 2013;63:1049-58.
- Lee HJ, Lee EK, Lee KJ, Hong SW, Yoon Y, Kim JS. Ectopic expression of neutrophil gelatinase-associated lipocalin suppresses the invasion and liver metastasis of colon cancer cells. *Int J Cancer*. 2006;118:2490–7.
- Leng X, Ding T, Lin H, Wang Y, Hu L, Hu J, Feig B, Zhang W, Pusztai L, Symmans WF, Wu Y, Arlinghaus RB. Inhibition of lipocalin 2 impairs breast tumorigenesis and metastasis. *Cancer Res*. 2009;69:8579–84.
- Leng X, Lin H, Ding T, Wang Y, Wu Y, Klumpp S, Sun T, Zhou Y, Monaco P, Belmont J, Aderem A, Akira S, Strong R, et al. Lipocalin 2 is required for BCR-ABL-induced tumorigenesis. *Oncogene*. 2008;27:6110–9.
- Li EM (1), Xu LY, Cai WJ, Xiong HQ, Shen ZY, Zeng Y. Functions of neutrophil gelatinase-associated lipocalin in the esophageal carcinoma cell line SHEEC. *Sheng Wu Hua Xue Yu Sheng Wu Wu Li Xue Bao (Shanghai)* 2003;35:247–54.

- Li EM (2), Xu LY, Xiong HQ, Cai WJ, Wu BL, Zhang C, Zhang YF, Lin Y, Shen ZY. Cloning and identification of 5'-untranslated region (UTR) and 3'-untranslated region of neutrophil gelatinase-associated lipocalin (NGAL) gene from esophageal carcinoma cell line SHEEC. *Ai Zheng*. 2003;22:143–7.
- Li SH, Hawthorne VS, Neal CL, Sanghera S, Xu J, Yang J, Guo H, Steeg PS, Yu D. Upregulation of neutrophil gelatinase-associated lipocalin by ErbB2 through nuclear factor-kappaB activation. *Cancer Res*. 2009;69:9163–8.
- Lim R, Ahmed N, Borregaard N, Riley C, Wafai R, Thompson EW, Quinn MA, Rice GE. Neutrophil gelatinase-associated lipocalin (NGAL) an early-screening biomarker for ovarian cancer: NGAL is associated with epidermal growth factor-induced epithelio-mesenchymal transition. *Int J Cancer*. 2007;120:2426–34.
- Lin CW, Tseng SW, Yang SF, Ko CP, Lin CH, Wei LH, Chien MH, Hsieh YS. Role of lipocalin 2 and its complex with matrix metalloproteinase-9 in oral cancer. *Oral Dis*. 2012;18:734–40.
- Lin J, Kamat A, Gu J, Chen M, Dinney CP, Forman MR, Wu X. Dietary intake of vegetables and fruits and the modification effects of GSTM1 and NAT2 genotypes on bladder cancer risk. *Cancer Epidemiol Biomarkers Prev*. 2009 Jul;18:2090-7.
- Liu MF, Jin T, Shen JH, Shen ZY, Zheng ZC, Zhang ZL, Xu LY, Li EM, Xu HX. NGAL and NGALR are frequently overexpressed in human gliomas and are associated with clinical prognosis. *J Neurooncol*. 2011;104:119–27.
- Martí J, Fuster J, Hotter G, Solà AM, Deulofeu R, Modolo MM, Loera MA, Ferrer J, Fondevila C, García-Valdecasas JC. Serum neutrophil gelatinase-associated lipocalin in patients with colorectal liver metastases: preliminary results of an exploratory prospective study. *Int J Biol Markers*. 2010;25:21–6.
- Martí J, Fuster J, Solà AM, Hotter G, Molina R, Pelegrina A, Ferrer J, Deulofeu R, Fondevila C, García-Valdecasas JC. Prognostic value of serum neutrophil gelatinase-associated lipocalin in metastatic and nonmetastatic colorectal cancer. *World J Surg*. 2013;37:1103–9.

- May WL, Johnson WD. A SAS Macro for the multivariate extension of the Kruskal-Wallis test including multiple comparisons: Randomization and  $\chi^2$  criteria. *Stat Soft Newsletter* 1997; 26:239-250.
- McLean MH, Thomson AJ, Murray GI, Fyfe N, Hold GL, El-Omar EM. Expression of neutrophil gelatinase-associated lipocalin in colorectal neoplastic progression: a marker of malignant potential? *Br J Cancer*. 2013;108:2537–41.
- Moniaux N, Chakraborty S, Yalniz M, Gonzalez J, Shostrom VK, Standop J, Lele SM, Ouellette M, Pour PM, Sasson AR, Brand RE, Hollingsworth MA, Jain M, et al. Early diagnosis of pancreatic cancer: neutrophil gelatinase-associated lipocalin as a marker of pancreatic intraepithelial neoplasia. *Br J Cancer*. 2008 May. 6;98:1540–7.
- Monier F, Mollier S, Guillot M, Rambeaud JJ, Morel F, Zaoui P. Urinary release of 72 and 92 kDa gelatinases, TIMPs, N-GAL and conventional prognostic factors in urothelial carcinomas. *Eur Urol*. 2002;42:356–63.
- Monier F, Surla A, Guillot M, Morel F. Gelatinase isoforms in urine from bladder cancer patients. *Clin Chim Acta*. 2000;299:11–23.
- Neilands JB. Siderophores: structure and function of microbial iron transport compounds. *J Biol Chem*. 1995;270:26723–26726.
- Nielsen BS, Borregaard N, Bundgaard JR, Timshel S, Sehested M, Kjeldsen L. Induction of NGAL synthesis in epithelial cells of human colorectal neoplasia and inflammatory bowel diseases. *Gut*. 1996;38:414–20.
- Nutt JE, Durkan JK, Lunec J. Matrix metalloproteinases (MMPs) in bladder cancer: the induction of MMP9 by epidermal growth factor and its detection in urine. *BJU Int* 2003;91:99-104.
- Okayama H, Kohno T, Ishii Y, Shimada Y, Shiraishi K, Iwakawa R, Furuta K, Tsuta K, Shibata T, Yamamoto S, Watanabe S, Sakamoto H, Kumamoto K, et al. Identification of genes upregulated in ALK-positive and EGFR/KRAS/ALK-negative lung adenocarcinomas. *Cancer Res*. 2012;72:100–11.

- Paley PJ. Ovarian cancer screening: are we making any progress? *Curr Opin Oncol.* 2001;13:399–402.
- Pei H, Li L, Fridley BL, Jenkins GD, Kalari KR, Lingle W, Petersen G, Lou Z, Wang L. FKBP51 affects cancer cell response to chemotherapy by negatively regulating Akt. *Cancer Cell.* 2009;16:259–66.
- Polesel J, Gheit T, Talamini R, Shahzad N, Lenardon O, Sylla B, La Vecchia C, Serraino D, Tommasino M, Franceschi S. Urinary human polyomavirus and papilloma virus infection and bladder cancer risk. *Br J Cancer* 2012; 106:222-226.
- Porta C, Paglino C, de Amici M, Quaglini S, Sacchi L, Imarisio I, Canipari C. Predictive value of baseline serum vascular endothelial growth factor and neutrophil gelatinase-associated lipocalin in advanced kidney cancer patients receiving sunitinib. *Kidney Int.* 2010;77:809–815.
- Provatopoulou X, Gounaris A, Kalogera E, Zagouri F, Flessas I, Goussetis E, Nonni A, Papassotiriou I, Zografos G. Circulating levels of matrix metalloproteinase-9 (MMP-9), neutrophil gelatinase-associated lipocalin (NGAL) and their complex MMP-9/NGAL in breast cancer disease. *BMC Cancer.* 2009;9:390.
- Rathmell WK, Godley PA. Recent updates in renal cell carcinoma. *Curr Opin Oncol.* 2010;22:250–256.
- Riker AI, Enkemann SA, Fodstad O, Liu S, Ren S, Morris C, Xi Y, Howell P, Metge B, Samant RS, Shevde LA, Li W, Eschrich S, et al. The gene expression profiles of primary and metastatic melanoma yields a transition point of tumor progression and metastasis. *BMC Med Genomics.* 2008;1:13.
- Roessler S, Jia HL, Budhu A, Forgues M, Ye QH, Lee JS, Thorgeirsson SS, Sun Z, Tang ZY, Qin LX, Wang XW. A unique metastasis gene signature enables prediction of tumor relapse in early-stage hepatocellular carcinoma patients. *Cancer Res.* 2010;70:10202–12.
- Rosser CJ, Ross S, Chang M, Dai Y, Mengual L, Zhang G, Kim J, Urquidi V, Alcaraz A, Goodison S. Multiplex protein signature for the detection of bladder cancer in voided urine samples. *J Urol* 2013;190:2257-62.

- Roy R, Louis G, Loughlin KR, Wiederschain D, Kilroy SM, Lamb CC, Zurakowski D, Moses MA. Tumor-specific urinary matrix metalloproteinase fingerprinting: identification of high molecular weight urinary matrix metalloproteinase species. *Clin Cancer Res.* 2008;14:6610–7.
- Sanchez-Carbayo M, Socci ND, Lozano J, Saint F, Cordon-Cardo C. Defining molecular profiles of poor outcome in patients with invasive bladder cancer using oligonucleotide microarrays. *J Clin Oncol.* 2006;24:778–89.
- Schlingemann J, Habtemichael N, Ittrich C, Toedt G, Kramer H, Hambek M, Knecht R, Lichter P, Stauber R, Hahn M. Patient-based cross-platform comparison of oligonucleotide microarray expression profiles. *Lab Invest.* 2005;85:1024–39.
- Scotto L, Narayan G, Nandula SV, Arias-Pulido H, Subramaniam S, Schneider A, Kaufmann AM, Wright JD, Pothuri B, Mansukhani M, Murty VV. Identification of copy number gain and overexpressed genes on chromosome arm 20q by an integrative genomic approach in cervical cancer: potential role in progression. *Genes Chromosomes Cancer.* 2008;47:755–65.
- Segara D, Biankin AV, Kench JG, Langusch CC, Dawson AC, Skalicky DA, Gotley DC, Coleman MJ, Sutherland RL, Henshall SM. Expression of HOXB2, a retinoic acid signaling target in pancreatic cancer and pancreatic intraepithelial neoplasia. *Clin Cancer Res.* 2005;11:3587–96.
- Sengupta S, den Boon JA, Chen IH, Newton MA, Dahl DB, Chen M, Cheng YJ, Westra WH, Chen CJ, Hildesheim A, Sugden B, Ahlquist P. Genome-wide expression profiling reveals EBV-associated inhibition of MHC class I expression in nasopharyngeal carcinoma. *Cancer Res.* 2006;66:7999–8006.
- Shay G, Lynch CC, Fingleton B. Moving targets: Emerging roles for MMPs in cancer progression and metastasis. *Matrix Biol.* 2015. 44-46C:200-206
- Shen ZZ, Zhao W, Gu J, Zhang ZQ, Yan L. Expression of matrix metalloproteinase-9 and its complex in the urine of breast cancer patients. *Zhonghua Wai Ke Za Zhi.* 2003;41:817–9.
- Shi H, Gu Y, Yang J, Xu L, Mi W, Yu W. Lipocalin 2 promotes lung metastasis of murine breast cancer cells. *J Exp Clin Cancer Res.* 2008;27:83.

- Shin SY, Kim JH, Baker A, Lim Y, Lee YH. Transcription factor Egr-1 is essential for maximal matrix metalloproteinase-9 transcription by tumor necrosis factor alpha. *Mol Cancer Res*. 2010 Apr;8(4):507-19.
- Sier CFM, Casetta G, Verhijen JH, Tizzani A, Agape V, Kos J, Blasi F, Hanemaaijer R. Enhanced urinary gelatinase activities (Matrix Metalloproteinases 2 and 9) are associated with early-stage bladder carcinoma: A comparison with clinically used tumor markers. *Clin Cancer Res* 2000;6:2333-40.
- Skrzypczak M, Goryca K, Rubel T, Paziewska A, Mikula M, Jarosz D, Pachlewski J, Oledzki J, Ostrowski J. Modeling oncogenic signaling in colon tumors by multidirectional analyses of microarray data directed for maximization of analytical reliability. *PLoS One*. 2010;5:e13091.
- Smith ER, Zurakowski D, Saad A, Scott RM, Moses MA. Urinary biomarkers predict brain tumor presence and response to therapy. *Clin Cancer Res*. 2008;14:2378–86.
- Stemmermann GN, Fenoglio-Preiser C. Gastric carcinoma distal to the cardia: a review of the epidemiological pathology of the precursors to a preventable cancer. *Pathology*. 2002;34:494–503.
- Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol*. 2001;17:463-516.
- Stoesz SP, Friedl A, Haag JD, Lindstrom MJ, Clark GM, Gould MN. Heterogeneous expression of the lipocalin NGAL in primary breast cancers. *Int J Cancer*. 1998;79:565–72.
- Su H, Hu N, Yang HH, Wang C, Takikita M, Wang QH, Giffen C, Clifford R, Hewitt SM, Shou JZ, Goldstein AM, Lee MP, Taylor PR. Global gene expression profiling and validation in esophageal squamous cell carcinoma and its association with clinical phenotypes. *Clin Cancer Res*. 2011;17:2955–66.
- Su LJ, Chang CW, Wu YC, Chen KC, Lin CJ, Liang SC, Lin CH, Whang-Peng J, Hsu SL, Chen CH, Huang CY. Selection of DDX5 as a novel internal control for Q-RT-PCR from microarray data using a block bootstrap re-sampling scheme. *BMC Genomics*. 2007;8:140.

- Syrjänen S, Naud P, Sarian L, Derchain S, Roteli-Martins C, Tatti S, Branca M, Erzen M, Hammes LS, Costa S, Longatto-Filho A, Syrjänen K. Up-regulation of lipocalin 2 is associated with high-risk human papillomavirus and grade of cervical lesion at baseline but does not predict outcomes of infections or incident cervical intraepithelial neoplasia. *Am J Clin Pathol.* 2010;134:50–9.
- Tang L, Zirpoli GR, Guru K, Moysich KB, Zhang Y, Ambrosone CB, McCann SE. Intake of cruciferous vegetables modifies bladder cancer survival. *Cancer Epidemiol Biomarkers Prev.* 2010 Jul;19(7):1806-11.
- Terris B, Blaveri E, Crnogorac-Jurcevic T, Jones M, Missiaglia E, Ruzniewski P, Sauvanet A, Lemoine NR. Characterization of gene expression profiles in intraductal papillary-mucinous tumors of the pancreas. *Am J Pathol.* 2002;160:1745–1754.
- Tong Z, Kunnumakkara AB, Wang H, Matsuo Y, Diagaradjane P, Harikumar KB, Ramachandran V, Sung B, Chakraborty A, Bresalier RS, Logsdon C, Aggarwal BB, Krishnan S, et al. Neutrophil gelatinase-associated lipocalin: a novel suppressor of invasion and angiogenesis in pancreatic cancer. *Cancer Res.* 2008;68:6100–8.
- Tsuji S, Midorikawa Y, Takahashi T, Yagi K, Takayama T, Yoshida K, Sugiyama Y, Aburatani H. Potential responders to FOLFOX therapy for colorectal cancer by Random Forests analysis. *Br J Cancer.* 2012;106:126–32.
- Tung MC, Hsieh SC, Yang SF, Cheng CW, Tsai RT, Wang SC, Huang MH, Hsieh YH. Knockdown of lipocalin-2 suppresses the growth and invasion of prostate cancer cells. *Prostate.* 2013;73:1281–90.
- Uemura N, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M, Taniyama K, Sasaki N, Schlemper RJ. *Helicobacter pylori* infection and the development of gastric cancer. *N Engl J Med.* 2001;345:784–9.
- Valk PJ, Verhaak RG, Beijten MA, Erpelinck CA, Barjesteh van Waalwijk van Doorn-Khosrovani S, Boer JM, Beverloo HB, Moorhouse MJ, van der Spek PJ, Löwenberg B, Delwel R. Prognostically useful gene-expression profiles in acute myeloid leukemia. *N Engl J Med.* 2004;350:1617–28.

- Vanaja DK, Chevillie JC, Iturria SJ, Young CY. Transcriptional silencing of zinc finger protein 185 identified by expression profiling is associated with prostate cancer progression. *Cancer Res.* 2003;63:3877–82.
- Vandooren J, Van den Steen PE, Opdenakker G. Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9): the next decade. *Crit Rev Biochem Mol Biol.* 2013;48:222-72.
- Varambally S, Yu J, Laxman B, Rhodes DR, Mehra R, Tomlins SA, Shah RB, Chandran U, Monzon FA, Becich MJ, Wei JT, Pienta KJ, Ghosh D, et al. Integrative genomic and proteomic analysis of prostate cancer reveals signatures of metastatic progression. *Cancer Cell.* 2005;8:393–406.
- Vasko V, Espinosa AV, Scouten W, He H, Auer H, Liyanarachchi S, Larin A, Savchenko V, Francis GL, de la Chapelle A, Saji M, Ringel MD. Gene expression and functional evidence of epithelial-to-mesenchymal transition in papillary thyroid carcinoma invasion. *Proc Natl Acad Sci U S A.* 2007;104:2803–8.
- Venkatesha S, Hanai J, Seth P, Karumanchi SA, Sukhatme VP. Lipocalin 2 antagonizes the proangiogenic action of ras in transformed cells. *Mol Cancer Res.* 2006;4:821–829.
- Villalva C, Sorel N, Bonnet ML, Guilhot J, Mayeur-Rousse C, Guilhot F, Chomel JC, Turhan AG. Neutrophil gelatinase-associated lipocalin expression in chronic myeloid leukemia. *Leuk Lymphoma.* 2008;49:984–8.
- Walker G, MacLeod K, Williams AR, Cameron DA, Smyth JF, Langdon SP. Estrogen-regulated gene expression predicts response to endocrine therapy in patients with ovarian cancer. *Gynecol Oncol.* 2007;106:461–8.
- Wang HJ, He XJ, Ma YY, Jiang XT, Xia YJ, Ye ZY, Zhao ZS, Tao HQ. Expressions of neutrophil gelatinase-associated lipocalin in gastric cancer: a potential biomarker for prognosis and an ancillary diagnostic test. *Anat Rec (Hoboken)* 2010;293:1855–63.
- Wenners AS, Mehta K, Loibl S, Park H, Mueller B, Arnold N, Hamann S, Weimer J, Ataseven B, Darb-Esfahani S, Schem C, Mundhenke C, Khandan F, et al. Neutrophil gelatinase-associated lipocalin (NGAL) predicts response to neoadjuvant chemotherapy and clinical outcome in primary human breast cancer. *PLoS One.* 2012;7:e45826.

- Wurmbach E, Chen YB, Khitrov G, Zhang W, Roayaie S, Schwartz M, Fiel I, Thung S, Mazzaferro V, Bruix J, Bottinger E, Friedman S, Waxman S, et al. Genome-wide molecular profiles of HCV-induced dysplasia and hepatocellular carcinoma. *Hepatology*. 2007;45:938–47.
- Xu L, Shen SS, Hoshida Y, Subramanian A, Ross K, Brunet JP, Wagner SN, Ramaswamy S, Mesirov JP, Hynes RO. Gene expression changes in an animal melanoma model correlate with aggressiveness of human melanoma metastases. *Mol Cancer Res*. 2008;6:760–9.
- Yan C, Boyd DD. Regulation of matrix metalloproteinase gene expression. *J Cell Physiol*. 2007; 211:19-26.
- Yan L, Borregaard N, Kjeldsen L, Moses MA. The high molecular weight urinary matrix metalloproteinase (MMP) activity is a complex of gelatinase B/MMP-9 and neutrophil gelatinase-associated lipocalin (NGAL). Modulation of MMP-9 activity by NGAL. *J Biol Chem*. 2001;276:37258–65.
- Yang J, Goetz D, Li JY, Wang W, Mori K, Setlik D, Du T, Erdjument-Bromage H, Tempst P, Strong R, Barasch J. An iron delivery pathway mediated by a lipocalin. *Mol Cell*. 2002;10:1045–56.
- Yang J, Moses MA. Lipocalin 2: a multifaceted modulator of human cancer. *Cell Cycle*. 2009;8:2347–52.
- Yang WC, Lin PM, Yang MY, Liu YC, Chang CS, Chou WC, Hsu JF, Huang CT, Cho SF, Yu WH, Lin SF. Higher lipocalin 2 expression may represent an independent favorable prognostic factor in cytogenetically normal acute myeloid leukemia. *Leuk Lymphoma*. 2013;54:1614–25.
- Yerlikaya G, Laml T, Elenskaia K, Hanzal E, Kölbl H, Umek W. Pain perception during outpatient cystoscopy: a prospective controlled study. *Eur J Obstet Gynecol* 2014;173:101-05.
- Yusenko MV, Kuiper RP, Boethe T, Ljungberg B, Van Kessel AG, Kovacs G. High-resolution DNA copy number and gene expression analyses distinguish chromophobe renal cell carcinomas and renal oncocytomas. *BMC Cancer*. 2009;9:152.

Zabron AA, Horneffer-van der Sluis VM, Wadsworth CA, Laird F, Gierula M, Thillainayagam AV, Vlavianos P, Westaby D, Taylor-Robinson SD, Edwards RJ, Khan SA. Elevated levels of neutrophil gelatinase-associated lipocalin in bile from patients with malignant pancreatobiliary disease. *Am J Gastroenterol.* 2011;106:1711–7.

Zhang H, Xu L, Xiao D, Xie J, Zeng H, Wang Z, Zhang X, Niu Y, Shen Z, Shen J, Wu X, Li E. Upregulation of neutrophil gelatinase-associated lipocalin in oesophageal squamous cell carcinoma: significant correlation with cell differentiation and tumour invasion. *J Clin Pathol.* 2007;60:555–61.

Zhang Y, Fan Y, Mei Z. NGAL and NGALR overexpression in human hepatocellular carcinoma toward a molecular prognostic classification. *Cancer Epidemiol.* 2012;36:e294–9.